# Taxonomy of Rhinolophus simplex Andersen, 1905 (Chiroptera : Rhinolophidae) in Nusa Tenggara and Maluku, Indonesia 

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#### Abstract

Rhinolophus simplex simplex Andersen, 1905 was collected for the first time from Bali, Nusa Penida, Moyo, Sangeang, Rinca, Flores, Lembata, Alor and Sumba islands. Additionally, specimens were collected from Lombok, Sumbawa. Other forms of R. simplex were collected from Timor, Savu, Roti, Semau and Kai Kecil islands. Rhinolophus simplex parvus Goodwin, 1979 is restricted to Timor Island; Rhinolophus simplex keyensis Peters, 1871 occurs on Kai Kecil Island. Rhinolophus simplex subsp. nov. is described on the basis of its morphology. It occurs on Savu, Roti and Semau islands.

Protein electrophoresis reveals that populations of Rhinolophus simplex show levels of genetic variation that are near the mammalian average. Genetic differentiation of populations is low.


## INTRODUCTION

Rhinolophus simplex was described by Andersen (1905) from Lombok I., Nusa Tenggara. Since then it has been considered a species by Corbet and Hill (1980, 1986, 1991); Honacki et al. (1982); van Strien (1986) and Tate and Archbold (1939). Koopman (1982), however, considered that R. simplex and the closely allied Maluku species: R. keyensis keyensis Peters, 1871; R. k. annectens Sanborn, 1939; R. truncatus Peters, 1871 and R. nanus Andersen, 1905 might well be subspecies of R. megaphyllus Gray, 1834. Hill (1992) supported this latter view and considered the above forms, along with $R$. robinsoni Andersen, 1918 and R. megaphyllus thaianus Hill, 1992 (Thailand) and R. klossi Andersen, 1918 (Malaya), as subspecies of R. megaphyllus.
Hill (1992) concluded that the major difference between the above forms was in the width of the sella, especially the base, and in the degree of inflation of the median anterior rostral swellings. He considered the forms from New Guinea and Australia and the forms thaianus, robinsoni and klossi from Thailand, have a wide sella and prominent, almost hemispherical narial swellings; in the Maluku forms the sella is a little narrower, but the narial swellings remain well developed; and in the Lesser Sunda island forms both sella and narial swellings are least developed.
Vertebrate surveys by the Western Australian Museum, in collaboration with the Museum Zoologicum Bogoriense, throughout Java, Lesser

Sunda islands and Maluku Tenggara, between 1987 and 1992, resulted in the collection of extensive series of specimens, closely related to $R$. simplex. Additionally a series of R. borneensis parvus Goodwin, 1979 (placed as a subspecies of $R$. celebensis by Hill 1992) was collected from Timor.
This paper reports on an examination of morphological and genetic variation among these specimens and on a comparison of this variation with a series of R. megaphyllus from Queensland, Australia. A small collection of $R$. borneensis importunus from Java are included for reference.

## MATERIALS AND METHODS

A total of 117 adult specimens was examined from a number of islands in Indonesia and Queensland, Australia (see Figure 1 for locality of specimens). They are listed in the "Specimens Examined" section. All these specimens are currently lodged in the Western Australian Museum (WAM). At the completion of this series of surveys half of all the WAM specimens, including the holotype, will be lodged in the Museum Zoologicum Bogoriense, Bogor.
Thirty two measurements of skull, dentary and dental characters and 18 of external characters (all in mm ) were recorded from adult specimens (see Figure 2, caption). The skull, dentary and dental characters were measured to an accuracy of 0.01 mm , while the external characters were measured to an accuracy of 0.1 mm .


Figure 1 Locality of Rhinolophus simplex and R. borneensis specimens used in this study. $\bullet$, Rhinolophus simplex simplex; $■$, R.s.parvus; $\star, R$. simplex keyensis; $\mathbf{\Delta}, R$. simplex subsp. nov.; and $\Delta$, Rhinolophus borneensis importunus.

Terminology used in the description of skull, dentary and dental (skull) characters and external characters follows Hill and Smith (1984). Pelage descriptions follow the colour terminology of Smithe (1975).
Adults were diagnosed as those specimens with basioccipital and sphenoid bones completely fused and epiphyseal swellings absent from metacarpal joints. Additionally two adult age classes were established based on extent of wear on $\mathrm{M}^{2}$ hypocone as follows: young adult, no wear or little wear such that the worn area is still elevated above the unworn hypocone basin; for adults, worn surface area of hypocone below level of unworn hypocone basin.
The effect of sex, adult age classes and taxion on skull, dental and external characters was investigated by stepwise multiple regressions on taxon, sex and age for five taxa. These were Rhinolophus megaphyllus (Queensland); R. borneensis importunus (Java); R. simplex simplex (Bali, Nusa Penida, Lombok, Sumbawa, Moyo, Sangeang, Rinca, Flores, Lembata, Alor and Sumba); R. simplex parvus (Timor) and R. simplex subsp. nov. (Savu, Roti and Semau). R. simplex keyensis was not included because the sample size was so small. Further, for the three R. simplex subspecies considered, the effect of sex, age and island on skull dental and external measurements was examined using multiple regressions. Examination of the residuals from regression analyses gave no indication of heteroscedasticity.

Canonical variate (discriminant) analysis (DFA) was computed on skull and external characters separately, with males and females combined, using the SPSS PC ${ }^{+}$program.

Cellogel electrophoresis of homogenised liver was used to investigate genetically determined protein variation using the techniques described in Richardson et al. (1986). This permitted the investigation of variation at 30 presumptive loci.

Genetic variation was assessed on 101 specimens, including some that were juvenile and not included in the morphometric analyses. The proteins scored, with Enzyme Commission Numbers and Locus Symbols in parenthesis, were: aconitate hydratase (E.C.4.2.1.3; Acon-1 \& Acon-2), adenosine deaminase (E.C.3.5.4.4; Ada), carbonate dehydratase (E.C.4.2.1.1; Ca), diaphorase (E.C.1.8.1.4; Dia), enolase (E.C.4.2.1.11; Enol), fructose-1, 6-diphosphatase (E.C.3.1.3.11; Fdp), fumarate hydratase (E.C.4.2.1.2; Fum), glucose-6phosphate dehydrogenase (E.C.1.1.1.49; G6pd), glyceraldehyde-3-phosphate dehydrogenase (E.C.1.2.1.12; Gapd), guanine deaminase (E.C.3.5.4.3; Gda), aspartate aminotransferase (E.C.2.6.1.1; Got-1 and Got-2), $\alpha$ glycerophosphate dehydrogenase (E.C.1.1.1.8; $\alpha \mathrm{Gpd}$ ), glucosephosphate isomerase (E.C.5.3.1.9; Gp-1), isocitrate dehydrogenase (E.C.1.1.1.42; Idh-1 and Idh-2), lactate dehydrogenase (E.C.1.1.1.27; $L d h-1$ and $L d h$ 2), malate dehydrogenase (E.C.1.1.1.37; Mdh-1 and Mdh-2), mannose-phosphate isomerase (E.C.5.3.1.8; Mpi ), purine nucleoside phosphorylase (E.C.2.4.2.1; $N p$ ), peptidase (E.C.3.4.13.11; Pep-A; E.C.3.4.11.4; Pep-B; E.C.3.4.13.11; Pep-C1 and E.C.3.4.13.9 Pep-D), 6-phosphogluconate dehydrogenase (E.C.1.1.44; 6Pgd), phosphoglucomutase (E.C.5.4.2.2; $\mathrm{Pg}^{\mathrm{m}}$ ), superoxide dismutase (E.C.1.15.1.1; Sod).

Chi-square was used to test for significance of contingency tables. Tables were reduced when more than a quarter of the cells had expected values less than 2 . When expected numbers were small after the tables were reduced to $2 \times 2$, exact probabilities were computed using twice the probability of the observed tail. Methods used to estimate heterozygosity within populations and genetic distances between populations were those of Nei (1978). These produce "unbiased" estimates. $F$-statistics were computed by the method of Weir and Cockerham (1984), which take into account


Figure 2 Skull and external measurements referred to in text and their recording points. GSL, greatest skull length; BB, braincase breadth; ZW, zygomatic width; MW, mastoid width; SBS, cranial height - excluding lambdoidal crest; RH , rostrum height $-\mathrm{M}^{1}$ alveoli to narial crest; RL , rostrum length - orbit anterior edge to maxillary anterior edge; IOB, minimum interorbital breadth; PIL, cranial length - junction of supraorbital and lambdoidal ridges to posteromost point of cranium; NIL, nasal inflation length - from above junction to nares; NIB, lateral narial inflation breadth; MSF, mesopterygoid fossa breadth; SW, sphenoid/pterygoid bridge basal breadth; SFB, sphenorbital fissure maximum breadth; CW, maximum cochlea breadth; CL, cochlea length; CCW, outside cochleae width; $\mathrm{M}^{3} \mathrm{M}^{3} \mathrm{~W}$, outside $\mathrm{M}^{3} \mathrm{M}^{3}$ width (cusp); $\mathrm{C}^{1} \mathrm{C}^{1} \mathrm{~W}$, outside $\mathrm{C}^{1} \mathrm{C}^{1}$ basal width (cusp); $\mathrm{C}^{1} \mathrm{~W}, \mathrm{C}^{1}$ width (cusp); $\mathrm{P}^{1} \mathrm{~W}$, first upper premolar width; $\mathrm{P}^{3} \mathrm{~L}$, last upper premolar length; $P^{3} W$, last upper premolar width; $M^{1} L, M^{1} W, M^{2} L, M^{2} W, M^{3} L$ and $M^{3} W$; first, second and third upper molar length and width, respectively; $\mathrm{C}^{1} \mathrm{M}^{3} \mathrm{~L}$, upper canine to $\mathrm{M}^{3}$ length (cusp); $\mathrm{I}_{1} \mathrm{M}_{3} \mathrm{~L}$, lower tooth row length (cusp); DL, dentary length - condyle to premaxilla anterior edge; SV, snout to vent length; TV, tail to vent length; EL, ear length; TIB, tibia length; PES, pes length; FA, forearm length; D2M, digit 2 metacarpal length; D3M, digit 3 metacarpal length; D3P1, digit 3 phalanx 1 length; D3P2, digit 3 phalanx 2 length; D4M, digit 4 metacarpal length; D4P1, digit 4 phalanx 1 length; D4P2, digit 4 phalanx 2 length; ALB, maximum anterior noseleaf breadth; BSL, basal sella length; BSB, maximum basal sella breadth; VSH, vertical sella height; and VSB, maximum vertical sella breadth.

Table 1 Measurements, in mm, (see Figure 2 caption for code to characters) for adult Rhinolophus megaphyllus, R. simplex simplex, R. s. parvus, R. simplex subsp. nov. and R. borneensis importunus. N, sample size; $\bar{X}$, mean; SD, standard deviation; min, minimum; max, maximum. (a) skull, dentary and dental characters and (b) external characters; males and females combined.

|  |  | GSL | BB | ZW | MW | SBS | RH | RL | IOB | PIL | NIL | NIB | MSF | SW | SFB | CW | CL |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| R. megaphyllus | $\underline{N}$ | 14 | 15 | 15 | 15 | 13 | 15 | 15 | 15 | 14 | 15 | 15 | 14 | 15 | 15 | 15 | 15 |
|  | $\bar{X}$ | 19.53 | 8.88 | 9.60 | 9.32 | 6.84 | 6.33 | 6.61 | 2.63 | 11.35 | 5.80 | 5.55 | 2.86 | 1.65 | 3.24 | 3.58 | 3.19 |
|  | SD | 0.43 | 0.18 | 0.24 | 0.19 | 0.25 | 0.27 | 0.20 | 0.10 | 0.30 | 0.31 | 0.19 | 0.16 | 0.12 | 0.16 | 0.10 | 0.10 |
|  | Min | 18.60 | 8.55 | 9.05 | 9.00 | 6.40 | 5.85 | 6.25 | 2.50 | 10.65 | 5.25 | 5.20 | 2.55 | 1.40 | 2.80 | 3.40 | 2.90 |
|  | Max | 20.00 | 9.15 | 10.00 | 9.55 | 7.25 | 6.70 | 6.90 | 2.85 | 11.75 | 6.25 | 5.80 | 3.10 | 1.85 | 3.50 | 3.75 | 3.35 |
| R. simplex simplex | N | 47 | 49 | 49 | 47 | 48 | 48 | 48 | 49 | 48 | 49 | 49 | 43 | 47 | 46 | 49 | 49 |
|  | $\bar{X}$ | 17.66 | 7.96 | 8.73 | 8.48 | 5.97 | 5.60 | 5.95 | 2.35 | 10.41 | 5.23 | 4.93 | 2.39 | 1.37 | 2.79 | 3.45 | 3.03 |
|  | SD | 0.42 | 0.20 | 0.25 | 0.17 | 0.32 | 0.18 | 0.27 | 0.12 | 0.32 | 0.21 | 0.17 | 0.12 | 0.10 | 0.15 | 0.13 | 0.08 |
|  | Min | 16.75 | 7.40 | 8.05 | 8.05 | 5.40 | 5.25 | 5.40 | 2.10 | 9.60 | 4.65 | 4.60 | 2.06 | 1.10 | 2.50 | 3.20 | 2.90 |
|  | Max | 18.45 | 8.45 | 9.15 | 8.80 | 6.80 | 6.10 | 6.50 | 2.60 | 11.00 | 5.65 | 5.50 | 2.60 | 1.60 | 3.10 | 3.75 | 3.20 |
| R. simplex keyensis | $\underline{N}$ | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
|  | $\bar{X}$ | 17.63 | 8.05 | 8.78 | 8.40 | 5.93 | 5.73 | 6.00 | 2.30 | 10.40 | 5.35 | 4.85 | 2.58 | 1.42 | 2.90 | 3.55 | 3.05 |
|  | SD | 0.18 | 0.21 | 0.18 | 0.07 | 0.32 | 0.25 | 0.14 | 0.07 | 0.57 | 0.64 | 0.28 | 0.25 | 0.04 | 0.07 | 0.0 | 0.07 |
|  | Min | 17.50 | 7.90 | 8.65 | 8.35 | 5.70 | 5.55 | 5.90 | 2.25 | 10.00 | 4.90 | 4.65 | 2.40 | 1.40 | 2.85 | 3.55 | 3.00 |
|  | Max | 17.75 | 8.20 | 8.90 | 8.45 | 6.15 | 5.90 | 6.10 | 2.35 | 10.80 | 5.80 | 5.05 | 2.75 | 1.45 | 2.95 | 3.55 | 3.10 |
| R. simplex parvus | $\underline{N}$ | $13$ | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 10 | 13 | 13 | 12 | $12$ |
|  | $\bar{X}$ | $16.87$ | 7.53 | 8.34 | 8.14 | 5.62 | 5.23 | 5.59 | 2.20 | $10.17$ | $4.57$ | 4.55 | $2.34$ | $1.40$ | $2.75$ | $3.28$ | $2.97$ |
|  | SD | 0.29 | 0.15 | 0.13 | 0.18 | 0.35 | 0.18 | 0.13 | 0.11 | 0.24 | 0.28 | 0.11 | 0.07 | 0.12 | 0.08 | 0.11 | 0.09 |
|  | Min | 16.50 | 7.20 | 8.08 | 7.79 | 5.10 | 5.00 | 5.40 | 2.04 | 9.75 | 4.20 | 4.35 | 2.20 | 1.16 | 2.60 | 3.10 | 2.85 |
|  | Max | 17.30 | 7.75 | 8.57 | 8.49 | 6.20 | 5.65 | 5.80 | 2.38 | 10.50 | 5.00 | 4.75 | 2.45 | 1.60 | 2.90 | 3.40 | 3.10 |
| $R$. simplex subsp. nov. | $\frac{\mathrm{N}}{\mathrm{x}}$ | 26 | 27 | 26 | 26 | 27 | 28 | 28 | 28 | 27 | 28 | 28 | 23 | 27 | 27 | 27 | 27 |
|  | $\bar{X}$ | 15.95 | 7.11 | 7.71 | 7.67 | 5.57 | 5.06 | 5.23 | 2.07 | 9.52 | 4.57 | 4.41 | 2.27 | 1.28 | 2.58 | 3.17 | 2.86 |
|  | SD | 0.50 | 0.29 | 0.34 | 0.26 | 0.35 | 0.23 | 0.23 | 0.11 | 0.33 | 0.20 | 0.17 | 0.14 | 0.07 | 0.11 | 0.09 | 0.11 |
|  | Min | 15.30 | 6.60 | 7.25 | 7.25 | 4.75 | 4.65 | 4.85 | 1.90 | 9.00 | 4.20 | 4.10 | 2.05 | 1.15 | 2.35 | 3.00 | 2.70 |
|  | Max | 17.10 | 7.65 | 8.40 | 8.20 | 6.40 | 5.55 | 5.70 | 2.30 | 10.25 | 4.95 | 4.80 | 2.50 | 1.40 | 2.80 | 3.40 | 3.15 |
| R. bormeensis importunus | $\underline{N}$ | 9 | 10 |  | 10 | 9 | 10 | 10 | 10 | $9$ | 10 | 9 | 10 | 9 | 8 | 9 | 10 |
|  | $\bar{X}$ | 19.2 | 8.67 | 9.72 | 9.08 | 6.43 | 6.10 | 6.41 | 2.34 | 11.55 | 5.07 | 5.26 | 2.74 | 1.59 | 2.89 | 3.54 | 3.06 |
|  | SD | 0.50 | 0.25 | 0.19 | 0.19 | 0.25 | 0.18 | 0.31 | 0.14 | 0.44 | 0.19 | 0.13 | 0.08 | 0.09 | 0.16 | 0.09 | 0.08 |
|  | Min | 18.50 | 8.20 | 9.45 | 8.85 | 5.90 | 5.75 | 5.85 | 2.10 | 10.50 | 4.75 | 5.05 | 2.60 | 1.40 | 2.70 | 3.45 | 2.90 |
|  | Max | 19.80 | 8.95 | 10.00 | 9.40 | 6.70 | 6.40 | 6.80 | 2.60 | 11.95 | 5.26 | 5.40 | 2.80 | 1.70 | 3.20 | 3.70 | 3.20 |


| Table 1 (continued) |  | CCW | $\mathbf{M}^{3} \mathbf{M}^{3} \mathbf{W}$ | $\mathrm{C}^{1} \mathrm{C}^{1} \mathrm{~W}$ | $\mathrm{C}^{1} \mathrm{~W}$ | $\mathbf{P}^{\mathbf{1}} \mathbf{W}$ | $\mathbf{P}^{\mathbf{3}} \mathbf{L}$ | $\mathrm{P}^{3} \mathrm{~W}$ | $\mathbf{M}^{1} \mathrm{~L}$ | $\mathrm{M}^{1} \mathbf{W}$ | $\mathrm{M}^{2} \mathrm{~L}$ | $\mathbf{M}^{\mathbf{2}} \mathbf{W}$ | $\mathbf{M}^{3} \mathrm{~L}$ | $\mathbf{M}^{3} \mathbf{W}$ | $\mathrm{C}^{1} \mathrm{M}^{3} \mathrm{~L}$ | $\mathrm{I}_{1} \mathrm{M}_{3} \mathrm{~L}$ | DL |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| R. megaphylus | N | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 14 |
|  | $\bar{\chi}$ | 4.90 | 7.16 | 7.86 | 1.26 | 0.49 | 1.08 | 1.51 | 1.57 | 1.55 | 1.62 | 1.66 | 1.55 | 1.57 | 7.34 | 8.52 | 12.65 |
|  | SD | 0.24 | 0.24 | 0.23 | 0.08 | 0.05 | 0.04 | 0.10 | 0.07 | 0.10 | 0.07 | 0.08 | 0.06 | 0.06 | 0.18 | 0.22 | 0.34 |
|  | Min | 4.40 | 6.70 | 7.30 | 1.05 | 0.40 | 1.00 | 1.30 | 1.50 | 1.35 | 1.50 | 1.55 | 1.50 | 1.45 | 7.05 | 8.05 | 12.05 |
|  | Max | 5.20 | 7.40 | 8.10 | 1.40 | 0.60 | 1.15 | 1.70 | 1.70 | 1.70 | 1.75 | 1.75 | 1.70 | 1.70 | 7.65 | 8.80 | 13.05 |
| R. simplex simplex | $\underline{N}$ | 48 | 48 | 48 | 48 | 48 | 48 | 48 | 49 | 49 | 49 | 49 | 48 | 48 | 47 | 47 | 5 |
|  | $\bar{\chi}$ | 4.40 | 6.34 | 7.48 | 1.08 | 0.42 | 0.98 | 1.36 | 1.42 | 1.37 | 1.41 | 1.43 | 1.31 | 1.38 | 6.65 | 7.80 | 11.64 |
|  | SD | 0.19 | 0.17 | 0.22 | 0.07 | 0.06 | 0.05 | 0.11 | 0.06 | 0.10 | 0.06 | 0.07 | 0.06 | 0.07 | 0.18 | 0.21 | 0.30 |
|  | Min | 3.90 | 5.80 | 6.85 | 0.95 | 0.30 | 0.90 | 1.10 | 1.30 | 1.20 | 1.25 | 1.25 | 1.20 | 1.15 | 6.30 | 7.40 | 11.00 |
|  | Max | 4.80 | 6.65 | 8.05 | 1.30 | 0.55 | 1.05 | 1.55 | 1.50 | 1.70 | 1.50 | 1.60 | 1.45 | 1.50 | 7.10 | 8.30 | 12.35 |
| R. simplex keyensis | N | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
|  | $\overline{\mathrm{X}}$ | 4.25 | 6.33 | 7.57 | 1.20 | 0.50 | 1.00 | 1.50 | 1.48 | 1.55 | 1.35 | 1.48 | 1.25 | 1.42 | 6.60 | 7.68 | 11.70 |
|  | SD | 0.07 | 0.18 | 0.04 | 0.0 | 0.0 | 0.0 | 0.0 | 0.04 | 0.0 | 0.0 | 0.04 | 0.07 | 0.04 | 0.14 | 0.25 | 0.28 |
|  | Min | 4.20 | 6.20 | 7.55 | 1.20 | 0.50 | 1.00 | 1.50 | 1.45 | 1.55 | 1.35 | 1.45 | 1.20 | 1.40 | 6.50 | 7.50 | 11.50 |
|  | Max | 4.30 | 6.45 | 7.60 | 1.20 | 0.50 | 1.00 | 1.50 | 1.50 | 1.55 | 1.35 | 1.50 | 1.30 | 1.45 | 6.70 | 7.85 | 11.90 |
| R. simplex parvus | N | 12 | 13 | 11 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 13 | 12 |
|  | $\bar{\chi}$ | 3.88 | 5.92 | 7.10 | 1.02 | 0.40 | 0.89 | 1.25 | 1.35 | 1.29 | 1.38 | 1.35 | 1.24 | 1.30 | 6.29 | 6.38 | 10.95 |
|  | SD | 0.15 | 0.15 | 0.13 | 0.08 | 0.06 | 0.05 | 0.10 | 0.07 | 0.09 | 0.06 | 0.05 | 0.05 | 0.08 | 0.17 | 0.17 | 0.29 |
|  | Min | 3.70 | 5.60 | 6.90 | 0.90 | 0.30 | 0.80 | 1.05 | 1.20 | 1.15 | 1.30 | 1.30 | 1.15 | 1.20 | 6.10 | 6.18 | 10.50 |
|  | Max | 4.10 | 6.10 | 7.35 | 1.15 | 0.55 | 1.00 | 1.40 | 1.45 | 1.40 | 1.50 | 1.45 | 1.30 | 1.40 | 6.60 | 6.68 | 11.25 |
| R. simplex subsp. nov. | N | 28 | 28 | 24 | 28 | 28 | 28 | 28 | 28 | 28 | 28 | 28 | 28 | 28 | 28 | 26 | 23 |
|  | $\bar{\chi}$ | 3.82 | 5.54 | 6.82 | 0.95 | 0.33 | 0.84 | 1.13 | 1.24 | 1.19 | 1.26 | 1.26 | 1.14 | 1.19 | 5.86 | 6.76 | 10.28 |
|  | SD | 0.21 | 0.19 | 0.18 | 0.07 | 0.07 | 0.04 | 0.09 | 0.05 | 0.07 | 0.07 | 0.08 | 0.06 | 0.07 | 0.26 | 0.26 | 0.30 |
|  | Min | 3.40 | 5.20 | 6.50 | 0.85 | 0.15 | 0.80 | 1.00 | 1.15 | 1.10 | 1.15 | 1.15 | 1.05 | 1.10 | 5.55 | 6.30 | 9.80 |
|  | Max | 4.25 | 5.90 | 7.10 | 1.10 | 0.45 | 0.95 | 1.35 | 1.35 | 1.35 | 1.40 | 1.45 | 1.30 | 1.40 | 6.40 | 7.30 | 10.90 |
| R. borneensis importunus | N | 10 | 10 | 9 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 10 | 9 |
|  | $\bar{\chi}$ | 4.80 | 6.97 | 7.87 | 1.30 | 0.51 | 1.09 | 1.53 | 1.64 | 1.53 | 1.64 | 1.60 | 1.48 | 1.54 | 7.31 | 8.56 | 12.63 |
|  | SD | 0.19 | 0.19 | 0.21 | 0.07 | 0.06 | 0.05 | 0.08 | 0.05 | 0.08 | 0.05 | 0.08 | 0.04 | 0.06 | 0.22 | 0.14 | 0.30 |
|  | Min | 4.50 | 6.60 | 7.50 | 1.15 | 0.40 | 1.00 | 1.45 | 1.55 | 1.45 | 1.55 | 1.50 | 1.40 | 1.45 | 6.90 | 8.35 | 12.10 |
|  | Max | 5.05 | 7.25 | 8.10 | 1.40 | 0.60 | 1.15 | 1.70 | 1.70 | 1.70 | 1.70 | 1.75 | 1.50 | 1.65 | 7.60 | 8.75 | 12.95 |


| Table 1 (continued) |  | SV | TV | EL | TIB | PES | FA | D2M | D3M | D3P1 | D3P2 | D4M | D4P1 | D4P2 | ALB | BSL | BSB | VSH | VSB |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| R. megaphyllus | N | 15 | 15 | 15 | 15 | 15 | 14 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 15 | 12 | 11 | 13 | 15 |
|  | $\overline{\mathrm{X}}$ | 47.2 | 24.1 | 19.4 | 19.1 | 8.3 | 46.9 | 35.1 | 32.9 | 13.5 | 19.9 | 34.5 | 10.2 | 12.1 | 9.0 | 2.5 | 3.1 | 4.4 | 2.2 |
|  | SD | 2.5 | 2.1 | 0.8 | 0.6 | 0.3 | 0.7 | 0.7 | 0.6 | 0.4 | 0.8 | 0.6 | 0.4 | 0.5 | 0.6 | 0.3 | 0.4 | 0.3 | 0.2 |
|  | Min | 42.6 | 20.9 | 17.6 | 17.5 | 7.4 | 45.8 | 33.3 | 31.9 | 12.7 | 18.1 | 33.5 | 9.8 | 11.3 | 7.7 | 2.0 | 2.7 | 3.7 | 1.8 |
|  | Max | 51.1 | 28.0 | 20.6 | 20.1 | 8.6 | 48.1 | 36.0 | 34.1 | 14.0 | 21.2 | 35.6 | 10.7 | 12.8 | 9.6 | 2.9 | 4.0 | 5.1 | 2.7 |
| R. simplex simplex | N | 49 | 49 | 49 | 48 | 49 | 48 | 48 | 49 | 49 | 49 | 49 | 49 | 49 | 49 | 39 | 38 | 40 | 47 |
|  | $\bar{\chi}$ | 43.2 | 21.1 | 17.7 | 18.2 | 7.6 | 41.9 | 31.4 | 29.5 | 12.3 | 19.1 | 30.3 | 8.8 | 11.8 | 8.1 | 2.3 | 2.5 | 3.9 | 2.0 |
|  | SD | 1.5 | 2.0 | 1.0 | 0.7 | 0.4 | 1.3 | 1.4 | 1.0 | 0.6 | 0.9 | 1.1 | 0.4 | 0.6 | 0.5 | 0.2 | 0.2 | 0.2 | 0.2 |
|  | Min | 39.9 | 18.1 | 14.4 | 16.0 | 6.1 | 39.0 | 28.7 | 27.1 | 11.0 | 17.1 | 27.5 | 7.5 | 10.5 | 7.2 | 1.8 | 2.1 | 3.2 | 1.7 |
|  | Max | 46.6 | 26.2 | 19.6 | 19.7 | 8.4 | 44.9 | 35.6 | 31.8 | 13.7 | 21.0 | 32.8 | 9.9 | 13.1 | 9.3 | 2.7 | 3.1 | 4.3 | 2.4 |
| R. simplex keyensis | $\underline{N}$ | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
|  | $\stackrel{\rightharpoonup}{\mathrm{x}}$ | 42.7 | 18.7 | 18.9 | 16.8 | 7.3 | 41.4 | 31.1 | 28.5 | 11.7 | 19.0 | 30.0 | 8.9 | $11.0$ | 9.2 | 2.7 | 2.8 | 4.5 | 2.4 |
|  | SD | 1.2 | 2.2 | 1.7 | 0.1 | 0.0 | 0.1 | 0.2 | 0.5 | 0.4 | 0.1 | 0.9 | 0.1 | 0.2 | 0.5 | 0.4 | 0.4 | 0.0 | 0.4 |
|  | Min | 41.8 | 17.2 | 17.7 | 16.7 | 7.3 | 41.4 | 31.0 | 28.2 | 11.4 | 19.0 | 29.4 | 8.8 | 10.9 | 8.8 | 2.4 | 2.6 | 4.5 | 2.1 |
|  | Max | 43.5 | 20.3 | 20.0 | 16.9 | 7.3 | 41.5 | 31.3 | 28.9 | 11.9 | 19.1 | 30.7 | 9.0 | 11.1 | 9.5 | 2.9 | 3.1 | 4.5 | 2.7 |
| R. simplex parvus | N | 13 | 12 | 13 | 13 | 13 | 13 | 12 | 12 | 13 | 12 | $12$ | 13 | 13 | 13 | 13 | 13 | 13 | $13$ |
|  | $\bar{\chi}$ | 37.7 | 20.1 | 16.1 | 17.3 | 7.7 | 40.1 | 29.9 | 27.9 | $11.6$ | $17.2$ | $28.7$ | 8.3 | $10.9$ | 7.4 | 1.9 | 2.5 | 3.4 | $1.9$ |
|  | SD | 1.2 | 2.2 | 0.4 | 0.6 | 0.5 | 0.8 | 0.7 | 1.0 | 0.4 | 0.5 | 0.9 | 0.3 | 0.4 | 0.3 | 0.2 | 0.1 | 0.3 | 0.1 |
|  | Min | 35.4 | 17.0 | 15.4 | 16.5 | 6.8 | 38.2 | 28.6 | 26.8 | 10.9 | 16.2 | 27.7 | 7.8 | 10.3 | 7.0 | 1.6 | 2.3 | 3.1 | 1.6 |
|  | Max | 39.7 | 23.8 | 16.6 | 18.5 | 8.3 | 41.3 | 31.1 | 30.2 | 12.3 | 18.0 | 30.5 | 8.5 | 11.8 | 7.8 | 2.2 | 2.7 | 4.0 | 2.0 |
| R. simplex subsp. nov. | N | 28 | 28 | $28$ | $28$ | 28 | 27 | 28 | 28 | 28 | 28 | 28 | 28 | 28 | 21 | 16 | 15 | 16 | 24 |
|  | $\bar{X}$ | 38.2 | 20.3 | $16.7$ | 15.2 | 6.4 | 37.4 | 27.6 | 26.3 | 10.3 | 16.2 | 26.7 | 7.9 | 9.8 | 7.6 | 2.3 | 2.4 | 3.6 | 1.9 |
|  | SD | 1.7 | 1.8 | 0.8 | 0.8 | 0.3 | 0.9 | 1.0 | 0.8 | 0.5 | 0.8 | 0.9 | 0.3 | 0.8 | 0.4 | 0.2 | 0.2 | 0.3 | 0.2 |
|  | Min | 35.6 | 18.0 | 15.0 | 14.0 | 5.9 | 35.8 | 26.1 | 24.8 | 9.5 | 14.7 | 24.6 | 7.5 | 8.4 | 6.8 | 2.0 | 2.2 | 3.3 | 1.6 |
|  | Max | 42.8 | 24.3 | 18.3 | 16.6 | 7.1 | 39.1 | 29.5 | 27.6 | 11.4 | 18.4 | 28.5 | 8.5 | 11.4 | 8.8 | 2.6 | 2.7 | 4.2 | 2.1 |
| R. borneensis importunus | N | 10 | 10 | 10 | 10 | 10 | 9 | 10 | 10 | 10 | 10 | 10 | 10 | $10$ | 10 | 9 | $9$ | $10$ | $10$ |
|  | $\bar{X}$ | 45.7 | 23.8 | 19.4 | 19.9 | 9.3 | 44.4 | 32.3 | 30.3 | 13.0 | 20.7 | 31.7 | 9.3 | 12.7 | 9.0 | 2.5 | 2.6 | 4.5 | 2.1 |
|  | SD | 2.4 | 2.3 | 0.54 | 0.9 | 0.4 | 1.0 | 1.1 | 0.9 | 0.5 | 1.0 | 1.2 | 0.4 | 0.6 | 0.3 | 0.2 | 0.2 | 0.3 | 0.1 |
|  | Min | 40.8 | 19.0 | 18.7 | 18.3 | 8.4 | 42.4 | 29.9 | 28.4 | 12.1 | 18.7 | 29.1 | 8.5 | 11.6 | 8.6 | 2.2 | 2.2 | 4.1 | 1.9 |
|  | Max | 48.1 | 26.9 | 20.4 | 21.1 | 9.7 | 45.5 | 33.6 | 31.3 | 13.6 | 22.1 | 33.3 | 10.0 | 13.6 | 9.4 | 3.0 | 2.8 | 5.0 | 2.2 |

sample size variation. Cluster analysis used the UPGMA method of Sneath and Sokal (1973). All statistical analyses were undertaken with Genstat 5 (Genstat 5 Committee 1987) and Biosys-1 (Swofford and Selander 1989).

## MORPHOLOGY: STATISTICS

## Univariate statistics

Mean, standard deviation, minimum and maximum values and sample size of each taxon are presented in Table 1 for (a) skull, dentary and dental characters and (b) external characters

## Multiple regressions

Multiple regressions were run for skull, dentary and dental characters and external characters on sex and adult age for five taxa ( $R$. borneensis, $R$. megaphyllus, R. simplex simplex, R. s. parvus and R. s. subsp. nov.). Additionally, multiple regression was run for the $R$. simplex group alone for skull and external characters on sex, adult age and island. In these analyses, islands with small sample size were omitted because either a sex or adult age category was absent. The results of these analyses are presented in Tables 2 and 3.
In the following discussions, because of the large number of interactions being tested, the level of significance was set at $\mathrm{P}<.01$.

## All 5 taxa - skulls

Sex. From Table 2a, eight characters (greatest skull length, GSL; braincase breadth, BB; rostrum height, RH; rostrum length, RL; outer cochlear width, CCW; upper maxillary tooth row length, $C^{1} M^{3} L$; lower tooth row length, $I^{1} M^{3} L$ and dentary length, DL) showed a significant relationship with sex alone ( $\mathrm{P}<0.000-\mathrm{P}=0.002$ ). Also there was a significant interaction for outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width for sex, age and taxon ( $\mathrm{P}=.009$ ). This interaction resulted from subadult females being smaller than subadult males in R. megaphyllus and females being slightly smaller than males in R. s. parvus and R. borneensis.
Age. Rostrum height, RH; showed a significant ( $\mathrm{P}=0.003$ ) relationship with age alone. Outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width also had a significant interaction between age, sex, and taxon ( $\mathrm{P}=0.009$ ), as discussed above.

Taxon. All skull characters, except $\mathrm{M}^{3}$ width, had a significant relationship ( $\mathrm{P}=<0.001$ ) with taxon. These relationships were consistent between the sex and age groupings; the only significant interaction was between sex, age and taxon ( $\mathrm{P}=0.009$ ) for outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width, as discussed above, clearly there is a considerable extent of morphological distinctness among these taxa.

## All 5 taxa-externals

Sex and age. There were significant relationships with snout to vent length, SV, both with sex alone
( $\mathrm{P}<0.001$ ) and age alone ( $\mathrm{P}<0.007$ ) and with interactions between sex and age and sex and taxon (Table 2b).
Taxon. All characters were significantly related to taxon ( $\mathrm{P}<0.001$ ), except basal sella length (BSL).

## R. simplex group - skulls

Sex. No character had a significant relationship with sex alone, although there was a significant interaction between age, sex and island ( $\mathrm{P}=0.005$ and $\mathrm{P}=0.002$, respectively) for braincase breadth BB and $\mathrm{M}^{2}$ width, $\mathrm{M}^{2} \mathrm{~W}$. For BB , this resulted from young adult females being larger than old adult females on Roti I., and males being larger than females on Moyo and Flores, whereas on other islands they were approximately the same size, For $\mathrm{M}^{2} \mathrm{~W}$, young adult males were larger than old adult males on Flores and young adult males were larger then young adult females on Savu I., whereas on other islands they were approximately the same size. Also, for $\mathrm{P}^{1}$ width, $\mathrm{P}^{1} \mathrm{~W}$, there was a significant interaction between sex and island ( $\mathrm{P}=0.008$ ) which resulted from females being larger than males on Savu I., whereas on other islands they were approximately the same size.

Age. No character had a significant relationship with age alone. But there were significant interactions for $B B$ and $M^{2} W$ between sex, age and island, as discussed above.
Island. All 30 characters had a significant relationship with island alone, most at $\mathrm{P} \ll 0.001$. These relationships were consistent for sex and age categories except for braincase breadth, $\mathrm{M}^{2}$ width, and $P^{1}$ width which had interactions between age and or sex and island, as discussed above.

Clearly, there was marked morphological differences among the island populations of $R$. simplex.

## R. simplex group - externals

Sex and age. The only significant relationship was the interaction between sex and island for ear length, EL (Table 3b). This resulted from the Rotinese sample where males had greater ear lengths than females, whereas on other islands they were subequal.

Island. All characters except ear length, EL; maximum anterior noseleaf breadth, ALB, and maximum basal sella breadth, BSB, were significant, most at $\mathrm{P}<0.001$. The only significant interaction was again between sex and island for ear length (Table 3b) as discussed above.

## Multivariate analyses

## 5 taxa analysis

Canonical variate (discriminant) analysis (DFA) was carried out on five taxa ( $R$. megaphyllus, $R$.

Table 2 Multiple regressions on taxon (Rhinolophus megaphyllus, R. simplex simplex, R. s parous, $R$. simplex subsp. nov. and $R$. borneensis importunus), sex and age for (a) skull, dentary and dental characters and (b) external characters. $F$ values are presented for the main effects and their interactions. Significance levels are ${ }^{*}, 0.05>p>0.01 ;{ }^{* *} 0.01>p>0.001$; and ${ }^{* * *} p<0.001$.

Table 2a

| Character | Main Effects |  |  | Interactions |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Sex | Age | Taxon | Sex. Age | Sex. Taxon | Age. <br> Taxon | Sex. Age. Taxon |
| GSL | ** |  | *** |  |  |  |  |
|  | 7.167 | 3.325 | 128.719 | 0.099 | 0.779 | 0.753 | 0.833 |
| BB | *** | * | *** |  |  |  |  |
|  | 11.472 | 4.116 | 155.075 | 0.235 | 0.778 | 0.416 | 0.402 |
| ZW |  |  | *** |  |  |  |  |
|  | 3.798 | 1.783 | 127.417 | 0.060 | 0.350 | 0.162 | 1.240 |
| MW | * |  | *** |  |  |  |  |
|  | 6.635 | 1.952 | 133.256 | 0.358 | 0.259 | 0.248 | 2.249 |
| SBS |  |  | *** |  |  |  |  |
|  | 1.445 | 0.487 | 26.914 | 0.009 | 0.785 | 0.767 | 0.511 |
| RH | *** | ** | ** |  |  |  |  |
|  | 11.602 | 9.752 | 64.575 | 0.280 | 0.843 | 0.710 | 1.640 |
| RL | ** |  | *** |  |  |  |  |
|  | 11.100 | 1.294 | 80.190 | 0.355 | 2.222 | 0.805 | 1.848 |
| 1 OB |  |  | ** |  |  |  |  |
|  | . 696 | . 904 | 25.446 | 0.932 | 0.856 | 0.982 | 0.898 |
| PIL |  |  | *** |  |  |  |  |
|  | 0.206 | 0.960 | 48.550 | 0.705 | 0.355 | 0.538 | 0.563 |
| NIL |  |  | *** |  |  |  |  |
|  | 1.717 | 0.314 | 34.673 | 2.103 | 0.160 | 0.165 | 1.253 |
| NIB | * |  | *** |  |  |  | * |
|  | 7.064 | 0.756 | 88.723 | 0.832 | 0.543 | 0.332 | 3.993 |
| sw |  |  | *** |  |  |  |  |
|  | 0.536 | 0.728 | 23.096 | 2.803 | 1.786 | 1.136 | 1.340 |
| SFB |  |  | *** |  |  |  |  |
|  | 0.908 | 0.930 | 29.172 | 2.802 | 0.879 | 0.472 | 0.560 |
| CW |  |  | *** |  |  |  |  |
|  | 0.177 | 0.106 | 20.200 | 0.839 | 0.308 | 0.666 | 0.360 |
| CL |  |  | ** |  |  |  |  |
|  | 0.288 | 0.248 | 23.949 | 2.588 | 0.644 | 0.443 | 0.864 |
| CCW | ** | * | *** |  |  |  | * |
|  | 9.626 | 6.572 | 63.086 | 0.125 | 0.753 | 0.353 | 4.005 |
| $\mathbf{M}^{3} \mathbf{M}^{3} \mathbf{W}$ |  |  | *** |  |  |  | ** |
|  | 3.631 | 0.778 | 156.608 | 0.230 | 1.114 | 0.745 | 5.091 |
| $C^{\prime} \mathrm{W}$ |  |  | *** |  |  |  |  |
|  | 0.152 | 0.000 | 31.014 | 3.093 | 1.208 | 1.130 | 1.907 |
| $\mathrm{P}^{\mathbf{1}} \mathrm{W}$ |  |  | *** |  |  |  | * |
|  | 0.062 | 0.599 | 13.425 | 1.258 | 1.741 | 2.065 | 3.510 |
| $\mathrm{P}^{3} \mathrm{~L}$ |  |  | *** | * |  |  |  |
|  | 0.444 | 0.079 | 59.942 | 6.123 | 0.322 | 1.179 | 2.854 |
| $\mathrm{P}^{3} \mathrm{~W}$ |  |  | *** |  |  |  |  |
|  | 0.375 | 0.003 | 29.514 | 2.202 | 0.365 | 0.670 | 0.931 |
| $\mathbf{M}^{\mathbf{1}} \mathrm{L}$ |  |  | *** |  |  |  |  |
|  | 0.223 | 0.785 | 68.903 | 0.628 | 0.240 | 0.589 | 1.007 |
| $\mathrm{M}^{\mathbf{1}} \mathbf{W}$ |  |  | *** |  |  |  |  |
|  | 0.105 | 0.619 | 21.441 | 0.904 | 0.610 | 0.257 | 0.743 |
| $\mathrm{M}^{2} \mathrm{~L}$ |  |  | *** |  |  |  |  |
|  | 0.244 | 0.047 | 63.528 | 0.104 | 0.894 | 1.121 | 0.626 |
| $\mathrm{M}^{2} \mathrm{~W}$ |  |  | **** |  |  |  |  |
|  | 3.792 | 1.159 | 41.020 | 1.011 | 1.367 | 0.459 | 1.168 |
| $\mathrm{M}^{3} \mathrm{~L}$ |  |  | *** | * |  |  |  |
|  | 0.746 | 0.038 | 59.181 | 4.298 | 0.802 | 0.901 | 1.700 |
| $\mathrm{M}^{3} \mathrm{~W}$ |  |  |  | * |  |  |  |
|  | 0.679 | 2.183 | 1.720 | 6.084 | 1.125 | 0.706 | 1.081 |
| $\mathrm{C}^{1} \mathrm{M}^{3} \mathrm{~L}$ | ** |  | ** |  |  |  | * |
|  | 8.473 | 1.459 | 116.743 | 0.000 | 0.305 | 0.637 | 3.323 |
| $\mathrm{I}_{1} \mathrm{M}_{3} \mathrm{~L}$ |  | * |  |  |  |  |  |
|  | 9.751 | 5.077 | 112.654 | 3.117 | 1.500 | 0.684 | 3.078 |
| DL | ** |  | *** |  |  |  |  |
|  | 10.616 | 2.973 | 144.502 | 0.015 | 1.092 | 1.165 | 2.409 |
| DEGREES OF FREEDOM | 1,71 | 1,71 | 4,71 | 1,71 | 4,71 | 4,71 | 2,71 |

Table 2b

borneensis importunus, R. simplex simplex, R. s. parvus and R. simplex subsp. nov.). Rhinolophus s. keyensis was included in this analyses but unallocated because of its small sample size ( $\mathrm{N}=2$ ). Both sex and age groups were combined. However, this analysis was carried out only after deleting nine skull characters shown in the multiple regression analysis to be significantly ( $\mathrm{P}<0.01$ ) influenced by sex or age (GSL, BB, RH, RL, CCW, $\mathrm{M}^{3} \mathrm{M}^{3} W$, $C^{1} \mathrm{M}^{3} \mathrm{~L}, \mathrm{I}^{1} \mathrm{M}^{3} \mathrm{~L}$, DL, see Table 2a). Also MSF, and $\mathrm{C}^{1} \mathrm{C}^{1} \mathrm{~B}$ were deleted because too many individuals had these values missing. The skull analyses used was with this reduced set of 21 characters.
Similarly, the DFA of external characters was run following deletion of snout to vent length, SVL, because there were significant ( $\mathrm{P}<0.01$ ) interactions between sex and age for this character (Table 3b).

Skulls. The DFA for the five taxa was first run using the reduced set of 21 characters and using islands as the a priori grouping. When these islands were grouped to represent the five above taxa the configuration of the taxon clusters in discriminate
function space was very similar. However, because the number of individuals in some taxa was less than the number of characters measured ( $R$. borneensis, 8 ; R. megaphyllus, 15) a reduced set of characters was used in the analysis. Of the 21 characters used in the DFA of the five taxa as the $a$ priori groups, five were chosen because they provided values that minimise Wilk's Lambda. The plots of the discriminant functions 1 to 3 , based on the reduced set of five characters (mastoid width, MW, supraorbital length, NIL; M ${ }^{1}$ length, $M^{1} L$; zygomatic width, ZW; and sphenorbital fissure width, SFB) produced very similar plots to the above analyses, and so only these are presented and discussed below.

The DFA produced three significant canonical functions. These three functions combined explained 99.9 percent of the variance (Table 4a) with function 1, 86.9 percent; function 2, 10.2 percent and function 3, 2.8 percent. A total of 95.3 percent of individuals were correctly classified to their appropriate taxon. Only five individuals were

Table 3 Multiple regressions for Rhinolophus simplex ( $R$. s. simplex, R.s. parvus and R.s. subsp. nov.) on island, sex, and age for (a) skull, dentary and dental characters and (b) external characters. F values are presented for the main effects and their interactions. Significance levels as for Table 2.

Table 3a


Table 3b

| Character | Main Effects |  |  | Interactions |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Sex | Age | Island | Sex. Age | Sex. Island | Age. Island | Sex. Age. Island |
| SV |  |  | *** |  |  |  |  |
|  | 0.176 | 0.069 | 28.275 | 0.150 | 0.520 | 0.694 | 2.084 |
| TV |  |  | * |  |  |  |  |
|  | 0.284 | 2.670 | 3.109 | 0.413 | 1.036 | 1.862 | 0.420 |
| EL |  |  |  |  | ** |  | * |
|  | 0.098 | 0.063 | 1.798 | 0.004 | 5.505 | 2.437 | 4.712 |
| TIB |  |  | *** |  |  |  |  |
|  | 0.005 | 0.083 | 41.150 | 0.926 | 0.281 | 0.365 | 1.472 |
| PES |  |  | *** |  |  |  |  |
|  | 0.116 | 0.040 | 12.590 | 0.904 | 0.917 | 0.912 | 0.818 |
| FA |  |  | *** |  |  |  |  |
|  | 0.543 | 1.035 | 24.474 | 1.183 | 0.838 | 0.931 | 3.275 |
| D2M |  |  | *** |  |  |  |  |
|  | 0.532 | 0.000 | 17.119 | 1.075 | 0.543 | 0.770 | 1.490 |
| D3M |  |  | *** |  |  |  |  |
|  | 0.072 | 1.232 | 14.872 | 1.342 | 0.547 | 0.679 | 0.249 |
| D3P1 |  |  | *** |  |  |  |  |
|  | 1.012 | 0.124 | 15.439 | 0.000 | 1.498 | 0.608 | 0.209 |
| D3P2 |  |  | *** |  |  |  |  |
|  | 0.799 | 1.046 | 17.561 | 0.199 | 0.880 | 0.426 | 0.002 |
| D4M |  |  | *** | * |  |  |  |
|  | 0.402 | 3.601 | 33.937 | 4.652 | 2.621 | 1.614 | 1.607 |
| D4P1 | * |  | *** |  |  |  |  |
|  | 4.457 | 0.012 | 13.284 | 1.016 | 2.557 | 0.866 | 0.019 |
| D4P2 |  |  | *** |  |  |  | * |
|  | 0.041 | 0.157 | 17.845 | 2.890 | 0.379 | 0.922 | 5.557 |
| ALB |  |  |  |  |  |  |  |
|  | 0.000 | 0.583 | 2.459 | 0.055 | 1.251 | 1.045 | 0.018 |
| BSL |  |  | ** |  |  |  |  |
|  | 0.094 | 0.165 | 4.530 | 1.108 | 0.073 | 1.021 | 1.099 |
| BSB |  |  |  |  |  |  | * |
|  | 0.932 | 0.007 | 0.699 | 1.916 | 0.533 | 1.302 | 5.935 |
| VSH |  |  | *** |  |  |  |  |
|  | 0.115 | 0.490 | 8.392 | 3.426 | 1.103 | 0.692 | 2.510 |
| VSB |  | * | * |  |  |  |  |
|  | 0.580 | 5.218 | 2.924 | 0.205 | 0.294 | 1.351 | 0.381 |
|  |  |  |  |  |  |  |  |
| FREEDOM | 1,23 | 1,23 | 5,23 | 1,23 | 3,23 | 4,23 | 1,23 |

incorrectly classified: Four R. simplex sp. nov. were classified as R. s. parvus and one R. s. simplex was classified as R. s. parvus.
The plot of functions 1 and 2 most clearly separates the taxa (Figure 3a) and plots of other combinations of functions 1-3 do not further clarify graphically this separation. From Figure 3a, the $R$. borneensis and R. megaphyllus clusters are clearly separate on function 2 and these two species clearly cluster separately from the R. simplex subspecies on function 1. Further, the R. simplex subspecies clusters, which partially overlap, separate on function 1. The unallocated Rhinolophus s. keyensis grouped closely with R. s. simplex.
The character loading most heavily ( $>0.5$ ) on function 1 and which is presumed an important discriminant between R. borneensis, R. megaphyllus and $R$. simplex, and among the $R$. simplex subspecies, was $\mathrm{M}^{1}$ length (Table 4a). The characters loading heavily ( $>0.5$ ) on function 2 and
presumed important in discriminating between $R$. borneensis and R. megaphyllus were supraorbital length, NIL, and zygomatic width, ZW (Table 4a).
Externals. The DFA for the five taxa was first run using the reduced set of 17 characters and using island as the a priori grouping. When these islands were grouped to represent the five above taxa the configuration of the taxon clusters in discriminant function space was very similar to that produced above. However, because the number of individuals in some taxa was less than the number of characters measured (e.g. R. borneensis, 9; R. megaphyllus, 12) fewer characters were used in the analysis. Five the 17 characters used in the DFA as the a priori groups were selected (forearm length, FA; pes length, PES; vertical sella height VSH; digit 4, phalanx 1 length, D4P2) for analysis because they provided values that minimise Wilk's Lambda. These five characters produced similar DFA plots to those from the 17 characters. Only the DFA


Figure 3 Canonical variate analysis grouped by taxon (Rhinolophus megaphyllus, $\square ;$ R. simplex simplex; R. s. parvus; $R$. s. subsp. nov.; and R. borneensis importunus) based on (a) skull, dentary and dental characters and (b) external characters, for functions 1 and 2. Other taxon symbols as for Figure 1 caption.

Table 4 Canonical variate function coefficients for the five taxa: Rhinolophus megaphyllus, R. simplex simplex, R. s parvus, R.s. subsp. nov. and R. borneensis parvus. R. s. keyensis unallocated. Standardised values, followed by (in brackets) unstandardised values. (a) skull and dental; (b) external characters.
Table 4a

| Character | Function 1 |  | Function 2 |  | Function 3 |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| MW | 0.3517 | $(1.7477)$ | 0.4891 | $(2.4302)$ | 0.6221 | $(3.0911)$ |
| NIL | 0.2224 | $(0.9304)$ | 0.7231 | $(3.0253)$ | -0.7308 | $(-3.0575)$ |
| M'L | 0.5114 | $(8.5400)$ | -0.4025 | $(-6.7206)$ | -0.1993 | $(-3.3275)$ |
| ZW | 0.3518 | $(1.3538)$ | -0.8787 | $(-3.3811)$ | -0.4380 | $(-1.6854)$ |
| SFB | 0.1374 | $(0.9804)$ | 0.4699 | $(3.3522)$ | 0.7509 | $(5.3568)$ |
| CONSTANT |  | -45.9212 |  | -6.4634 |  | -6.3208 |
| VARIATION | 86.9 |  | 10.2 |  | 2.8 |  |
| EXPLAINED (\%) |  |  |  |  |  |  |

Table 4b

| Character | Function 1 |  | Function 2 |  | Function 3 |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| FA | 0.6108 | $(0.5706)$ | -0.1597 | $(-0.1492)$ | -0.1565 | $(-0.1462)$ |
| PES | 0.2590 | $(0.7415)$ | 1.0000 | $(2.8640)$ | -0.4652 | $(-1.3324)$ |
| VSH | 0.3455 | $(1.2204)$ | -0.2507 | $(-0.8854)$ | 0.2965 | $(1.0471)$ |
| D4P2 | -0.1158 | $(-0.1892)$ | 0.2766 | $(0.4519)$ | 1.1051 | $(1.8054)$ |
| D4M | 0.3276 | $(0.3474)$ | -0.6520 | $(-0.6915)$ | -0.1803 | $(-0.1912)$ |
| CONSTANT |  | -42.6952 |  | 3.4166 |  | -2.7095 |
| VARIATION | 77.3 |  | 16.6 |  | 4.1 |  |
| EXPLAINED (\%) |  |  |  |  |  |  |

based on this reduced set of five characters are presented and discussed below.

The DFA produced four significant canonical functions. These functions combined explained 100 percent of the variance (Table 4 b ) with function 1, 77.3 percent; function $2,16.6$ percent; function 3 , 4.1 percent and function $4,2.0$ percent. A total of 95.6 percent of individuals was correctly classified to their appropriate taxon. Four specimens were incorrectly classified. One R.s. subsp. nov. was classified as R.s. parvus, one R.s. parvus was classified as R.s. subsp. nov. and two R.s. simplex were classified as R.s. parous.

The plot of functions 1 and 2 and functions 1 and 3 most clearly separates the taxa (Figure 3b) and plots of other combinations of functions $1-4$ does not further clarify graphically this separation. From Figure 3b, R. borneensis, R. megaphyllus, R.s. simplex and R.s. subsp. nov. cluster separately on function 1 with R.s. parvus also clustering separately from $R$. megaphyllus and $R$. borneensis on this function.

Additionally $R$. megaphyllus clusters separately from both R.s. parvus and R. borneensis on function 2. The unallocated $R$. simplex keyensis specimens cluster close to R. s. simplex.

The character loading most heavily ( $>0.6$ ) on function 1, which is presumed to be a most important discriminant between most of these taxon, was forearm length (Table 4b). the characters loading heavily ( $>0.6$ ) on function 2 , and presumed important discriminants between $R$. megaphyllus and both R.s. parous and R. borneensis
were pes length, PES, and digit 4 metacarpal length, D4M (Table 4b).

## R. simplex analysis

A DFA was carried out on the three subspecies of R. simplex (R.s. simplex, R.s. parvus, and R. s. subsp. nov.) after combining both sex and age groups. This analysis was carried out for skulls, dentary and dental characters only after deleting three skull characters shown by the multiple regression analysis to be significantly ( $\mathrm{P}<0.01$ ) influenced by sex or age (BB, $\mathrm{P}^{1} \mathrm{~W}$, and $\mathrm{M}^{2} \mathrm{~W}$, see Table 3a). Also, MSF and $C^{1} C^{1} W$ were deleted because too many individuals had these values missing. The skull analyses was then run with the reduced set of 25 characters.

Similarly the DFA of external characters was run after deletion of ear length from the analysis because there was a significant ( $\mathrm{P}<0.01$ ) interaction between sex and island for this character (Table 3b).

Skulls. The DFA for the R. simplex subspecies was first run using the reduced set of 25 characters and using islands as the a priori groupings. When these islands were then grouped to represent the three taxa R. s. simplex, R. s. parvus and R. s. subsp. nov. (R. s. keyensis unallocated), the configuration of the taxon clusters in discriminant function space was similar to that produced above. However, as the number of characters was considerably larger than the number of individuals in one taxon group ( $R$. simplex parous, 12) a reduced set of five characters


Figure 4 Canonical variate analysis grouped by Rhinolophus subspecies (R. s. simplex, R. s. parvus and R.s. subsp. nov.) based on (a) skull, dentary and dental characters and (b) external characters, for functions 1 and 2. Islands symbols are as follows: a, Sumbawa; b, Bali; f, Flores; g, Sangeang; i, Rinca; k, Kai Kecil; I, Lombok; m, Moyo; o, Alor; p, Nusa Penida; r, Roti; s, Semau; t, Timor; u, Sumba; w, Savu.

Table 5 Canonical variate functions coefficients for the three subspecies of Rhinolophus simplex (R. s. simplex, R. s. parvus, R. s. subsp. nov.). Rhinolophus s. keyensis unallocated. Standardised values, followed by (in brackets) unstandardised values (a) skull and dental; (b) external characters.
Table 5a

| Character | Function 1 |  | Function 2 |  |
| :---: | :---: | :---: | :---: | :---: |
| $\mathrm{I}_{1} \mathrm{M}_{3} \mathrm{~L}$ | 1.2760 | $(5.8790)$ | -0.9938 | (-4.5787) |
| $\mathrm{C}^{1} \mathrm{M}^{3} \mathrm{~L}$ | $-0.8948$ | $(-4.4169)$ | 1.0576 | (5.2205) |
| $\mathrm{M}^{3} \mathrm{M}^{3} \mathrm{~W}$ | 0.1955 | (1.1858) | 0.8975 | (5.4438) |
| CB | 0.4427 | (3.8282) | 0.2631 | (2.2755) |
| NIB | 0.2770 | (1.6894) | -0.4586 | (-2.7968) |
| CONSTANT |  | -42.6169 |  | -27.1418 |
| VARIATION EXPLAINED (\%) | 75.4 |  | 24.6 |  |

Table 5b

|  | Function 1 |  | Function 2 |  |
| :--- | :--- | :--- | :--- | :--- |
| TIB | 0.4414 | $(0.6271)$ | -0.4069 | $(-0.5781)$ |
| SV | 0.9602 | $(0.6010)$ | 0.6795 | $(0.4506)$ |
| PES | 0.3907 | $(1.1571)$ | -0.8443 | $(-2.5005)$ |
| D4P1 | 0.4708 | $(1.3476)$ | 0.4592 | $(1.3145)$ |
| VSH | 0.6294 | $(3.3753)$ | 0.0508 | $(0.2722)$ |
| TV | -0.5209 | $(-0.2470)$ | 0.1439 | $(0.0682)$ |
| BSB | -0.5409 | $(-2.5337)$ | 0.3179 | $(1.4891)$ |
| CONSTANT |  | -50.4099 |  | -6.9463 |
| VARIATION | 85.4 | 14.6 |  |  |
| EXPLAINED (\%) |  |  |  |  |

was selected on the basis that they provided values that minimise Wilk's lambda. The plots of the discriminant function 1 and 2 based on this reduced set of five characters (lower, tooth row length, $\mathrm{I}_{1} \mathrm{M}_{3} \mathrm{~L}$; upper maxillary tooth row length, $C^{1} \mathrm{M}^{3} \mathrm{~L}$; outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width, $\mathrm{M}^{3} \mathrm{M}^{3} \mathrm{~W}$; cranium breadth, CB; and nasal inflation breadth, NIB) produced very similar plots to the above analyses, and so only these are presented and discussed below.
The DFA produced two significant canonical functions. Function 1 explained 75.4 percent of the variance and function $2,24.6$ percent (Table 5a). A total of 100 percent of individuals were correctly classified to their appropriate subspecies. The plot of function 1 and 2 (Figure 4a) clearly separates the subspecies: simplex, parvus and subsp. nov. with the unallocated keyensis grouping with simplex. Function 1 separates all three allocated subspecies clusters and function 2 partially separates $R$. s. simplex from both R.s parous and R. s. subsp. nov. and completely separates the R. s. parvus and R.s. subsp. nov. clusters.
The characters loading most heavily ( $>0.8$ ) on function 1 and presumed important discriminants between the three allocated subspecies, were lower tooth row length, $\mathrm{I}_{1} \mathrm{M}_{3} \mathrm{~L}$, and upper maxillary tooth row length, $\mathrm{C}^{1} \mathrm{M}^{3} \mathrm{~L}$ (Table 5a). Characters loading most heavily on function $2(>0.8)$ and presumed particularly important in discriminating between R. s. parvus and R. s subsp. nov. included, in
addition to the above two characters, outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width, $\mathrm{M}^{3} \mathrm{M}^{3} \mathrm{~W}$ (Table 5a).

Externals. The DFA for the R. simplex subspecies was first run using the reduced set of 17 characters and using islands as the a priori groupings. When these islands were then grouped to represent the three allocated R. simplex subspecies (see above), the configuration of the taxon clusters in discriminant function space was similar to that produced above. However, because the number of characters was larger than the number of individuals in one taxon group ( $R$. simplex parvus, 12) a reduced set of seven characters was selected (tibia length, TIB; snout to vent length, SV; pes length, PES; digit 4, phalanx 1 length, D4P1; vertical sella breadth, VSB; tail to vent length, TV; basal sella breadth, BSB) using the method for skulls above; this produced similar DFA plots to those produced using the 17 characters. Only the DFA based on this set of seven characters are presented and discussed below.

The DFA produced two significant canonical functions. Function 1 explained 85.4 percent of the variance and function $2,14.6$ percent (Table 5 b). A total of 100 percent of individuals were correctly classified to their appropriate subspecies. The plot of functions 1 and 2 (Figure 4b) clearly separates the subspecies, with the unallocated keyensis again grouping with R. s. simplex. Function 1 separates all three subspecies clusters and function 2 separates R.s. parvus and R.s. subsp. nov.


Figure 5 Dendrogram computed by UPGMA from the genetic distance (Nei standard distance, unbiased) between island populations of Rhinolophus simplex and R. borneensis from Java.

The characters loading most heavily ( $>0.6$ ) on function 1 and presumed important discriminants between R.s. simplex and both R. s. parvus and R.s. subsp. nov. are snout to vent length, SV and vertical sella height, VSH (Table 5b). Characters loading most heavily on function $2(>0.6)$ and presumed important discriminants between R.s. parous and R.s. subsp. nov. are snout to vent length and pes length, PES (Table 5b).

## GENETICS

The gene frequencies are presented in Table 6. Fifteen of the 30 loci scored showed variation within islands. The genotype frequencies for all occurrences of polymorphism within an island did not differ significantly from the Hardy-Weinberg expectations. Mean heterozygosity levels are presented at the bottom of Table 6. They fall within the usual range observed for mammalian populations (Nevo, Beiles and Ben-Shlomo 1984).
Much of the variation within Rhinolophus simplex was due to inter-island differences. F-statistics revealed four loci that had $F_{S T}$ greater than 0.1. These were Acon-2 (0.83), Idh-2 (0.63), Pep-D (0.16) and $6 \mathrm{Pgd}(0.22)$. The mean $F_{S T}$ over all loci was 0.42 with bootstrapped $95 \%$ confidence limits of 0.09 and 0.67 . However, for most loci, there was little or no variability within or between islands and the unbiased Nei genetic distances between islands was low, being generally less than 0.04 (Table 7). These distances, together with those estimated from a sample of 18 individuals of $R$. borneensis
from Java were subjected to cluster analysis and the resultant dendrogram is presented in Figure 5. This dendrogram reveals the integrity of $R$. simplex as a species distinct from R. borneensis.

## SYSTEMATICS

Rhinolophus simplex simplex Andersen, 1905
Rhinolophus simplex Andersen, K., 1905: 76, Pl. 3.

## Holotype

British Museum No. 97.4.18.4, adult female, in alcohol, collected June 1896 by A. Everett.

## Type locality

Lombok I., Nusa Tenggara, altitude 2500 ft (ca. 830 m ).

## Diagnosis

Rhinolophus s. simplex differs from both $R$. simplex parvus and $R$. simplex subsp. nov. in averaging larger in all skull, dental and dentary measurements, except for the posterior width of the sphenoid/pterygoid bridge; SW, and external measurements, except pes length, PES, and basal sella length, BSL, (Tables 1a, b). It differs almost absolutely from R.s subsp. nov. in tibia length and forearm length (see Table 1). $\mathrm{I}_{1} \mathrm{M}_{3}$ longer relative to outside cochleae width, nasal inflation breadth, $\mathrm{C}^{1} \mathrm{M}^{3}$ length and outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width (Figures 6, 7a, b, c, respectively). It differs from R.s. parous in having snout to vent length longer relative to pes


Figure 6 Plot of lower tooth row length, $\mathrm{I}_{1} \mathrm{M}_{3^{\prime}}$ versus cochlea breadth, CW, for Rhinolophus simplex subspecies, $R$. megaphyllus ( $\square$ ) and R. borneensis importunus. Other taxon symbols as for Figure 1.
length (Figure 8).
It is approximately the same size as R. s. keyensis. For example: greatest skull length 17.66 (16.7518.45) v. 17.63 (17.50-17.25), zygomatic width 8.73 (8.05-9.15) v. $8.78(8.65-8.90)$ and forearm length 41.9 (39.0-44.9) v. 41.4 (41.4-41.5)]. It differs from keyensis in having a smaller sella, with vertical sella height absolutely shorter 3.9 (3.2-4.3) v. 4.5 (4.54.5 ) and vertical sella breadth averaging narrower 2.0 (1.7-2.4) v. 2.4 (2.1-2.7).

## Description

## Skull and dentition

Rhinolophus s. simplex is larger than R. s. parvus and R. s. subsp. nov. (Table 1). Specimens show some variation in overall size and shape, but most of this variation may be found in a single population, for example, Moyo I. This can be appreciated by the wide distribution of the values for Moyo I. for the functions 1 and 2 scores from the DFA within the R. s. simplex cluster (Figure 4a). This is also true for those characters that were not quantified such as the size and shape of the anterior rostral swellings and the juxtaposition of the upper and lower vestigial premolar with the other premolars.

The junction of the supraorbital ridges is usually behind the mid-point of the orbital cavity but occasionally it is anterior to that point. The median anterior rostral swellings also vary considerably is size and extent of inflation in relation to the supraorbital length (Figure 9); some individuals, from throughout the range of $R$. s. simplex approximate the large size and inflation of the two individuals of $R$. s. keyensis from the eastern Kai Kecil I. The size and extent of crowding of the vestigial upper and lower premolar was extremely variable. Usually the anterior upper premolar was in contact with the canine but its contact with the posterior premolar was extremely variable. The lower vestigial premolar varied from being located almost in the tooth row between the anterior and posterior premolar, sometimes not in solid contact with these adjacent premolars, to being totally extruded such that the anterior and posterior premolars are in contact.

The anterior basisphenoid has a marked depression or pit which is much lower than the surface of the basioccipital. This is quite different from the shape of this region in $R$. megaphyllus ignifer where the basioccipital surface runs smoothly into a shallow basisphenoid groove.

The posterior palate margin terminates at $\mathrm{M}^{2}$ mid

Table 6 Allele frequencies, mean heterozygosity and sample sizes in island populations of Rhinolophus simplex and R. borneensis from Java. A dash indicates the allele was not detected. The mean heterozygosity and its standard error, and mean number of individuals ( N ) per locus are shown at the bottom of the table. No variation was detected at the following loci: Acon-1, Ca, Enol, Fdp, Fum, Gapd, G6pd, Got-1, Gp-1, Idh-1, Idh-2, Mdh-1, Mdh-2, Np and Pep-C1.

| LOCUS | GENE | BALI | N.PENIDA | ISLAND LOMBOK | SUMBAWA | MOYO | SANGEANG | RINCA |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Acon-2 | A | - | - | - | - | - | - | - |
|  | B | - | - | - | - | - |  |  |
|  | C | - | - | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
|  | D | 1.00 | 1.00 | - | - | - | - | - |
|  | E | - |  | - | - | - | - | - |
| Ada | A | - | - | - | - | - | - | - |
|  | B | - | - | - | 0.10 | - | - | - |
|  | C | 1.00 | 1.00 | 1.00 | 0.90 | 1.00 | 1.00 | 1.00 |
|  |  |  | - | - | - | - | - | - |
| Dia | A | - | - | - | 0.10 | 0.05 | $\overline{1}$ | - |
|  | B | 1.00 | 1.00 | 1.00 | 0.90 | 0.95 | 1.00 | 1.00 |
|  | C | - | - | - | - | - | - | - |
|  | D | - | - | - | - | - | - | - |
|  | E | - | - | - | - | - | - | - |
| Gda |  |  |  |  |  |  |  |  |
|  | B | - | - | $-$ |  |  |  | $-$ |
|  |  |  |  |  |  |  |  | 1.00 |
|  | A | - | - | - | - | - | - | 0.12 |
|  | B | $\overline{-}$ | 1 | 1 |  | 1 | 100 | - |
|  | C | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.88 |
| Got-2 |  |  | - | - | - | - | - | - |
|  | B | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
|  | $\mathrm{C}$ | - | - | - | - | - | - | - |
|  |  |  |  |  |  |  |  |  |
| Idh-2 | A | $\cdots$ | $\bar{\square}$ | - | $\overline{-}$ | 100 | 1.00 |  |
|  | B | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
|  | C | - | - | - | - | - | - | - |
| Ldh-1 | A | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.88 |
|  | B | - | - | 1.0 | - | - | - | 0.12 |
| Mpi | A | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |  | $1.00$ |
|  | B | - | - | - | - |  | - | $-$ |
| Pep-A | A | - | - | - | - | $\overline{-}$ | $\overline{-}$ | $\overline{100}$ |
|  | B | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
|  | C | - | - |  | - | - | - | - |
|  | D | - | - | - | - | - | - | - |
| Pep-B | $\mathrm{A}$ | - | - | - |  | 0.05 |  | - |
|  | B | - | $-$ | - | 0.10 | 0.05 | - | - |
|  | C | - | - |  |  |  |  | $\overline{100}$ |
|  | D | 1.00 | 1.00 | 1.00 | 0.80 | 0.90 | 1.00 | 1.00 |
|  | $\underset{\text { E }}{\text { E }}$ | - | - | - | $\overline{0.10}$ | - | - | - |
|  | F | - | - | - | 0.10 | - | - | - |
| Pep-D | A | - | - | - |  |  |  |  |
|  | ${ }^{\text {B }}$ | $\overline{1.00}$ | $\overline{1.00}$ | $\overline{1.00}$ | $\overline{0.80}$ | 0.30 0.60 | $\overline{1.00}$ | $\overline{1.00}$ |
|  | D | 1.00 | - | - | - | - | - | - |
|  | E | - | - | - | - | - | - | - |
|  | F | - |  | - | 0.20 | 0.10 | - | - |
|  | G | - | - | - | - | - | - | - |
| 6Pgd | A | 0.44 0.50 | 0.07 0.93 | 0.25 0.75 | 0.50 0.50 | 0.50 0.50 | 0.50 0.50 | $\begin{aligned} & 0.12 \\ & 0.88 \end{aligned}$ |
|  | $\stackrel{\text { C }}{ }$ | - | - | - | - | - | - | - |
|  | D | 0.06 | - | - | - | - | - | - |
|  | E | - | - | - | - | - | - | - |
| Pgm |  | $1.00$ | $1.00$ | $1.00$ | $1.00$ | $1.00$ | $1.00$ | $1.00$ |
|  | B |  |  |  |  |  |  |  |
| Sod |  |  |  |  |  |  |  |  |
|  | B | $1.00$ | $1.00$ | $1.00$ | $1.00$ | $1.00$ | $1.00$ | 1.00 |
|  |  | $0.019$ | $0.005$ | $0.017$ | $0.056$ | $0.046$ | $0.033$ | $0.025$ |
| Standard error |  | $0.019$ | $0.005$ | $0.017$ | $0.025$ | 0.026 | $0.033$ | $0.014$ |
| N per locus |  | 9 | 7 | 2 | 5 | 10 | 1 | 4 |

Table 6 (continued)

| SUMBA | FLORES | Lembata | ALOR | ISLAND TIMOR | SEMAU | ROTI | SAVU | JAVA |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| - | - | - | -- | - | - | 0.04 | - | 1.00 |
| - | - | - | - | - | 0.12 | - | - | - |
| 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.88 | 0.96 | - | - |
| - | - | - | - | - | - | - | 0.25 | - |
| - | - | - | - | - | - | - | 0.75 | - |
| - | - | - | - | - | - | - | - | 0.08 |
| - | - | - | - | - | - | - | - | 0.17 |
| 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.50 |
| - | - | - | - | - | - | - | - | 0.25 |
| - | - | - | - | - | - | - | - | - |
| 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.96 | 0.80 | 0.36 |
| - | - | - | - | - | - | - | - | 0.64 |
| - | - | - | - | - | - | 0.04 | - | - |
| - | - | - | - | - | - | - | 0.20 | - |
| - | - | - | 0.04 | - | - | - | - | - |
| $\overline{1.00}$ | 1.00 | $\overline{100}$ | -0.96 | - 100 | $\cdots$ | 1.00 | $\cdots$ | 1.00 |
| 1.00 | 1.00 | 1.00 | 0.96 | 1.00 | 1.00 | 1.00 | 1.00 | - |
| - | - | 0.17 | 0.05 | - | - | - | - | - |
| $\overline{1.00}$ | 1.00 | - 0.83 | $\overline{0.96}$ | $-$ | - 100 | - 100 | - | 0.97 |
| 1.00 | 1.00 | 0.83 | 0.96 | 1.00 | 1.00 | 1.00 | 1.00 | 0.03 |
| - | - | - | 0.04 | - | - | - | - | - |
| 1.00 | 1.00 | 1.00 | 0.96 | 1.00 | 1.00 | 1.00 | 0.92 | 0.94 |
| - | - | - | - | - | - | - | 0.08 | - |
| - | - | - | - | - | - | - | - | 0.06 |
| - | 0.25 | 0.33 | 0.73 | - | - | - | - | - |
| 1.00 | 0.75 | 0.67 | 0.23 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
| - | - | - | 0.04 | - | - | - | - | - |
| 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 |
| - | - | - | - | - | - | - | - | - |
| 1.00 | 1.00 | 1.00 | 0.86 | 1.00 | 1.00 | 0.96 | 0.98 | 0.75 |
| - | - | - | 0.14 | - | - | 0.04 | 0.02 | 0.25 |
| - | - | - | 0.04 | - | - | - | 0.02 | - |
| 0.92 | 1.00 | 0.83 | 0.96 | 1.00 | 1.00 | 1.00 | 0.98 | 0.09 |
| - | - | - | - | - | - | - | - | 0.03 |
| 0.08 | - | 0.17 | - | - | - | - | - | - |
| - | - | - | - | - | - | - | - | - |
| - | - | - | - | - | - | - | - | - |
| $-$ | - | - | $\cdots$ | - | - | - | - | 0.94 |
| 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | - |
| - | - | - | - | - | - | - | - | 0.06 |
| - | - | - | - | - | - | - | - | - |
| 0.08 | - | - | - | - | 0.17 | - | - | - |
| - | - | - | - | - | - | - | - | - |
| 0.92 | 0.75 | 1.00 | 0.46 | 0.92 | 0.66 | 0.73 | 1.00 | 1.00 |
| - | - | - | 0.09 | -- | - | - | - | - |
| - | - | - | 0.09 | - | - | - | - | - |
| - | 0.25 | - | 0.27 | 0.08 | 0.17 | 0.27 | - | - |
| - | - | - | 0.09 | - | - | - | - | - |
| 0.08 | 0.25 | 0.17 | - | 0.42 | 0.25 | - | - | - |
| 0.84 | 0.75 | 0.83 | 1.00 | 0.58 | 0.75 | 1.00 | 1.00 | 0.94 |
| 0.08 | - | - | - | - | - | - | - | - |
| - | - | - | - | - | - | - | - | - |
| - | - | - | - | - | - | - | - | 0.06 |
| 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.98 | 0.97 |
| - | - | - | - | - | - | - | 0.02 | 0.03 |
| - | - | - | - | - | 0.12 | - | - | - |
| 1.00 | 1.00 | 1.00 | 1.00 | 1.00 | 0.88 | 1.00 | 1.00 | 1.00 |
| 0.022 | 0.050 | 0.051 | 0.059 | 0.023 | 0.051 | 0.023 | 0.034 | 0.073 |
| 0.013 | 0.028 | 0.025 | 0.029 | 0.018 | 0.026 | 0.014 | 0.017 | 0.029 |
| 6 | 2 | 3 | 11 | 6 | 4 | 11 | 20 | 18 |





Outer $M^{\mathbf{3}} \mathrm{M}^{\mathbf{3}}$ width
Figure 7 Plot of lower tooth row length, $\mathrm{I}_{1} \mathrm{M}_{3}$, versus (a) nasal inflation breadth, NIB; (b) upper maxillary tooth row length, $\mathrm{C}^{1} \mathrm{M}^{3}$; and (c) outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width, $\mathrm{M}^{3} \mathrm{M}^{3} \mathrm{~W}$; for subspecies of Rhinolophus simplex; subspecies symbols as for Figure 1, caption.
point; occasionally there is a slight median postpalatal protuberance but more usually this is a smooth U shape; premaxilla posteriorly terminates at $\mathrm{P}^{3}$ mid point; sphenoidal/pterygoid bridge obscures anterior sphenorbital fissure when viewed from the ventral aspect; incisors weak, bilobed; $\mathrm{M}^{1-2}$ hypocone well developed.

## Externals

The external characters are similar to those described by Andersen (1905) for Rhinolophus simplex. The anterior noseleaf is moderately wide (8.2) with an obvious anteromedian notch. The vertical sella is longer than the basal sella (4.0 v . 2.3), smoothly curved at the apex and not noticeably constricted. Ears are moderately long (17.8), outer apical margin reasonably concave.

## Pelage and skin colour

Dorsal pelage of most specimens Fuscous, which is colour of distal one-third of hairs, the basal part of which is Drab. Ventral surface pure Drab.

Occasional specimens of both sex paler, with dorsal surface Russet, which is colour of distal onethird of hairs, the basal part of which is Cream Color. Ventral surface pure Fawn Color. Wing and ears Fuscous.

## Baculum

The shape of bacula among Rhinolophus simplex largely differs in the extent of the incision in the posteroventral basal margin (Figure 10, Table 8), which ranges from deeply incised in WAM M38252 (Bali I). to slightly concave in WAM M30249 (Sumba I). All these basal types occur in R. s simplex.

## Distribution

Many islands in the Inner Banda Arc: Bali, Nusa Penida, Lombok, Sumbawa, Moyo, Sangeang, Komodo, Rinca, Flores, Lembata, Alor, Wetar islands and Sumba Island in the outer Banda Arc. Its ability to occupy new habitats, such as defence tunnels build by Japanese during the Second World War, suggest that it is a good colonist and that it probably exists on many other islands in this region.

Rhinolophus simplex keyensis Peters, 1871
Rhinolophus megaphyllus keyensis Peters, W., 1871.

## Types

Cotypes, Mus. Berol. No. 3240 and No. 3291.

## Type locality

Key-Inseln (= Kai Islands).


Figure 8 Plot of snout to vent length versus pes length for subspecies of R. simplex. Subspecies codes as for Figure 1.

## Diagnosis

Rhinolophus s. keyensis differs from R. s. simplex as diagnosed earlier for this subspecies.

It differs from R. s. parvus in averaging larger in all skull, dentary and most external characters (see Table 1). With the following characters it is absolutely larger: ear length, forearm length, maximum anterior noseleaf breadth, basal sella length, vertical sella height, vertical sella breadth, greatest skull length, rostrum length and $\mathrm{M}^{1}$ width (see Table 1).
It differs from $R$. s. amiri subsp. nov. in averaging larger in most skull, dentary, dental and external characters (see Table 1). With a number of characters it is absolutely larger, for example: forearm length, vertical sella height, greatest skull length, zygomatic width, cranial width and $\mathrm{C}^{1} \mathrm{M}^{3}$ length (see Table 1).

## Description

Apart from differences described in the diagnosis of R. s. simplex, R. s. keyensis is very similar to that species. In the two specimens of $R$. s. keyensis available to us, the juncture of the supraorbital ridge is just posterior to the orbital cavity mid point, supraorbital length greater than nasal inflation breadth (Figure 9); median anterior rostral swellings inflated; anterior upper premolar in contact with both canine and posterior premolar;
lower vestigial premolar extruded from toothrow but still in contact with adjacent premolars - in WAM 42642 the anterior and posterior premolars are not in contact, whereas in WAM M42643 they are in solid contact.

The form of the basiphenoid, palate and dentition is similar to R. s. simpex.

## Externals

The external characters similar to R. s. simplex but anterior noseleaf wider ( $9.2 v .8 .1$ ). The vertical sella taller ( $4.5 v .3 .9$ ) and wider (2.4 v. 2.0) with a slightly wider mid part than $R$. s. simplex, smoothly rounded at apex.

Pelage and skin colour and baculum
As described for R. s. simplex.

## Distribution

Kai Kecil I.

## Remarks

This form is only weakly separated from R.s. simplex; the subspecific distinction resting solely on the shape and size of its sella. In all other characters it appears to be very close to $R$. s. simplex, as attested to by its closeness to that subspecies in discriminant function space (Figures


Figure 9 Plot of nasal inflation breadth, NIB, versus supraorbital length, NIL, for the taxa Rhinolophus simplex simplex, R. s. parvus, R. s. subsp. nov. R. megaphyllus (ם) and R. borneensis parvus. The oblique broken line joins values where NIB = NIL. Other taxon symbols as for Figure 1, caption. Following Hill (1992), those forms with values generally below the line should be R. megaphyllus while those generally above the line are other species in the 'ferrumequinum' group.

4a, b). Future collections from Tanimbar and Wetar Is may help clarify its taxonomic status with respect to R. s. simplex.

## Rhinolophus simplex parous Goodwin, 1979

Rhinolophus borneensis parvus Goodwin, 1979: 102105.

## Holotype

American Museum, Natural History No. 237766, adult male, skin and skull, collected 27 March 1968.

## Type locality

Lia Hoo Cave, nr Fatu Maca village, 11 km S Baucau, Timor, altitude ca. 550 m .

## Diagnosis

Rhinolophus simplex parvus differs from Rhinolophus s. simplex and R. s. keyensis as diagnosed earlier for these subspecies.
Differs from Rhinolophus simplex subsp. nov. in averaging larger (but with measurements overlapping) in all skull and dentary characters except supraorbital length and lower tooth row
length (Table 1a). Also the relationship between lower tooth row length and : nasal inflation breadth, $\mathrm{C}^{1} \mathrm{M}^{3}$ length and outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width differs (Figures 7a,b,c). General body measurements also average larger except those related to facial foliations: maximum noseleaf breadth, basal sella length, vertical sella height, maximum vertical sella breadth.

## Description

Apart from differences described in the diagnoses of R. s. simplex, R. s. parvus is very similar to that subspecies. The skull, however, tends to have the juncture of supraorbital and lambdoidal ridges level with orbital cavity mid point ( $36 \%$ ), just posterior ( $36 \%$ ) or well posterior from that point ( $28 \%$ ); anterior sphenoid/ pterygoid bridge tends to obscure more of sphenorbital sinus when viewed from ventral aspect.

Pelage and skin colour also differs slightly. Dorsal pelage Cinnamon Brown, which is colour of distal one-third of hairs, basal part of which is Drab. Ventral surface pure Fawn Color. Wings Grayish Brown, Ears Fawn Color.

## Distribution

Timor I. only.

## Rhinolophus simplex amiri Kitchener subsp. nov.

## Holotype

Museum Zoologicum Bogoriense (MZB) No. 15901 (field number WAM M3519; adult female; weight 3.6 gm ; carcase fixed in 10 percent formalin, preserved in 75 percent ethanol; skull separate; liver in ultrafreeze; collected by D.J. Kitchener on 26 September 1990.

## Type locality

Kampung Wawarae, Desa Menia, Savu I., Nusa Tenggara, Indonesia ( $10^{\circ} 29^{\prime} \mathrm{S}, 121^{\circ} 55^{\prime} \mathrm{E}$ ), collected by hand, from shallow limestone cave, in open woodland, 500 m from sea, altitude 20 m .

## Paratypes

Listed in "Specimens examined" section.

## Diagnosis

Rhinolophus simplex amiri differs from R. s. simplex, R. s. keyensis and R. s. parvus as described in the earlier diagnoses of these subspecies.

## Description

Apart from the differences described in the diagnosis and description of R. s. simplex and R. s. parous, $R$. s. amiri is very similar to R. s. parous. The colour of pelage is, however, variable, particularly the Savu specimens. Roti specimens have pelage similar in colour to the usual dark phase of R.s. simplex. Semau specimens are similar to $R$. s. parous.
Specimens from Savu are distinct and a much paler colour. The dorsal surface has hairs with the basal part Cream Color; on the rump this is tipped with Fuscous which is the dominant colour; on the neck, shoulders and upper back the Fuscous tipping is sparse allowing the Cream Color to appear to provide a vague paler mantle. The chest
and throat pure Cream Color; the abdomen darker because this Cream Color base tipped with Drab.

## Distribution

Semau, Roti, and Savu I., Nusa Tenggara Timur, Indonesia

## Etymology

Named after Dr M. Amir, Director, Balitbang Zoologi, LIPI, Indonesia, in recognition for his support of this collaborative research project between 1987 and 1993.

## Remarks

Rhinolophus simplex (R. s. simplex, R. s. parvus, $R$. s. amiri and R. s. keyensis) differ from R. megaphyllus ignifer in being generally smaller in most skull, dental, dentary and external measurements (see Table 1a, b). For example, greatest skull length 15.30-18.45 v. 18.60-20.00; braincase breadth 6.608.45 v. 8.55-9.15; zygomatic width 7.25-9.15 v. $9.05-10.00$; nasal inflation breadth $4.10-5.20$ v. 5.205.80; mastoid width 7.25-8.80 v. 9.00-9.55; outer $\mathrm{M}^{3} \mathrm{M}^{3}$ width $5.20-6.65$ v. 6.70-7.40; $\mathrm{C}^{1} \mathrm{M}^{3}$ length 5.55-7.10 v. 7.05-7.65; forearm length 35.8-44.9 v. 45.8-48.1. The anterior basisphenoid has a marked depression or pit, rather than a shallow groove that runs smoothly into the basioccipital surface. It also differs in having $I_{1} \mathrm{M}_{3}$ length smaller relative to outer cochleae width (Figure 6) and forearm length longer relative to pes length (Figure 11). Baculum base with ventral bifurcated arms much shorter relative to posterior margin of dorsal surface of base (Figure 10).

## DISCUSSION

Hill (1992) lists R. truncatus Peters, 1871 and R. nanus Andersen, 1905 from Bacan I., N. Maluku and Goram I., C. Maluku as a subspecies of $R$. megaphyllus. We have not been able to examine specimens of these forms. However, descriptions of them, along with measurements in Andersen (1905:84), suggest that R. truncatus is more allied to


Figure 10 Range of bacula size and shape in Rhinolophus simplex, compared to R. megaphyllus. (a) R. s. simplex (WAM 30249, Sumba); (b) R.s. subsp. nov (WAM M35374, Roti); (c) R. s. simplex (WAM M38252, Bali) and (d) R. megaphyllus (WAM M29979, Queensland).

Table 7 Nei's unbiased genetic distance between Rhinolophus simplex populations and R. borneensis from Java.

| NUSA PENIDA | 0.005 |  |  |  |  |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| LOMBOK | 0.034 | 0.033 |  |  |  |  |  |
| SUMBAWA | 0.035 | 0.041 | 0.000 |  |  |  |  |
| MOYO | 0.038 | 0.045 | 0.004 | 0.000 | 0.004 |  |  |
| SANGEANG | 0.034 | 0.041 | 0.000 | 0.000 | 0.008 | 0.004 |  |
| RINCA | 0.038 | 0.034 | 0.000 | 0.004 | 0.007 | 0.005 | 0.000 |
| SUMBA | 0.038 | 0.034 | 0.000 | 0.004 | 0.000 | 0.000 | 0.000 |
| FLORES | 0.034 | 0.033 | 0.000 | 0.000 | 0.009 | 0.005 | 0.000 |
| LEMBATA | 0.039 | 0.036 | 0.000 | 0.005 | 0.031 | 0.035 | 0.026 |
| ALOR | 0.070 | 0.062 | 0.026 | 0.030 | 0.002 | 0.000 | 0.002 |
| TIMOR | 0.033 | 0.038 | 0.000 | 0.000 | 0.001 | 0.002 | 0.000 |
| SEMAU | 0.032 | 0.031 | 0.000 | 0.000 | 0.010 | 0.011 | 0.002 |
| ROTI | 0.043 | 0.035 | 0.002 | 0.007 | 0.038 |  |  |
| SAVU | 0.028 | 0.020 | 0.029 | 0.038 | 0.042 | 0.038 | 0.030 |
| JAVA | 0.171 | 0.161 | 0.162 | 0.163 | 0.174 | 0.172 | 0.159 |
|  | BALI | NUSA P. | LOMBOK | SUMBAWA | MOYO | SANGEANG | RINCA |

R. megaphyllus whereas the narrower cranium and nasal swellings and overall small size of $R$. nanus are more suggestive of R. simplex. However, J.E. Hill (pers. comm.) informed us that the holotype of R. nanus is very old and has a damaged skull. Consequently, he considers that the narrowing of the skull of R. nanus may well be an artefact because otherwise the skull is very like that of $R$. truncatus.

Hill (1992:100) separated the R. megaphyllus species complex (in which he includes R. s. simplex and R. s. keyensis) from other IndoMalayan species in the ferrumequinum group principally on the supraorbital crests combining at a point behind the centre of the orbital cavity; such that the supraorbital length (from junction of crests to nares) is much greater than rather than slightly greater than or equal to the width across the anterior lateral rostral swellings (in our terminology NIL NIB) and the supraorbital depression being larger than it is wide. The association by Goodwin (1979) and Hill (1992) of the form parous with R. borneensis and R. celebensis, respectively, depended largely on the supraorbital crests of parous merging anterior to the mid point of the orbital cavity. However, in the sample of parvus available to us this junction point was variable; sometimes it was level with the point, or just behind or well behind it (an R. simplex character). Also the supraorbital length is frequently much greater than the width across the outer lateral rostral swellings (see also Figure 9). Clearly in the form parvus (and amiri) this latter character is too variable for it to be usefully diagnostic in terms of the association of parvus, although it appears to hold true for $R$. borneensis and generally so for R. megaphyllus and R. s. simplex. Goodwin (1979:104) further considered that $R$. simplex differed from parous in being larger
overall and in having "dentition (that) is somewhat more primitive. The vestigial premolars in both upper and lower jaws are generally not as crowded, but there is some individual variation in this condition" Further, "the sella of simplex is slightly constricted and the connecting process is not as prominent". In the specimens available to us there was considerable variation in the extent of crowding in both the upper and lower vestigial premolar, particularly the lower. The lower premolar in both parous and R. s. simplex varied in its position from almost being in the toothrow to being completely extruded such that this first and second premolars were in contact. Further we can find no consistent difference between parous and $R$. s. simplex in the shape of either the sella or the connecting process.
We associate the form parvus with R. simplex rather than with $R$. borneensis, as suggested by Goodwin (1979) or with R. celebensis as considered by Hill (1992). This is because of its morphological closeness to $R$. simplex and because, as discussed above, the characters used by these authors to diagnose it from R. simplex cannot be substantiated by us. It is also relevant here that our electrophoretic study, using 30 loci, concluded that there was little or no detectable genetic difference between R. s. simplex, R. s. parous and R. s. amiri. For example the population of R. s. parvus (Timor) is not genetically differentiated from several $R$. s. simplex populations (Sangeang, Sumbawa), while two populations of R. s. amiri (Roti and Semau) are closer genetically to the majority of the R. s. simplex populations than they are to the third population of R. s. amiri (Savu). The significance of the apparent clusters within R. simplex based on the Nei genetic distance metric is tenuous because it is the product of gene frequency variation at just one or two loci. Thus Alor differentiates due to

Table 7 (continued)

| 0.000 |  |  |  |  |  |  |  |  |
| :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- | :--- |
| 0.000 | 0.000 | 0.010 |  |  |  |  |  |  |
| 0.025 | 0.006 | 0.003 | 0.029 |  |  |  |  |  |
| 0.002 | 0.000 | 0.002 | 0.021 | 0.000 |  |  |  |  |
| 0.000 | 0.000 | 0.004 | 0.020 | 0.006 | 0.000 |  |  |  |
| 0.002 | 0.000 | 0.032 | 0.057 | 0.035 | 0.028 | 0.030 |  |  |
| 0.030 | 0.030 | 0.150 | 0.193 | 0.169 | 0.164 | 0.160 | 0.151 |  |
| 0.157 | 0.166 | LEORES | LEMBATA | ALOR | TIMOR | SEMAU | ROTI | SAVU |
| SUMBA |  |  |  |  |  |  |  |  |

variability at two loci, $I d h-2$ and Pep-D, while the Bali-Nusa Penida-Savu cluster is largely due to Acon-2 allele frequency differences.
Hill (1983) considered that the form parvus was very similar to $R$. madurensis Andersen, 1918 from Madura I., a view supported by Bergmans and van Bree (1986) who considered parvus synonymous with $R$. celebensis madurensis. We have been unable to examine specimens of the form madurensis, but if
parvus is indeed synonymous with $R$. celebensis, then it brings into question the distinction between other species in the ferrumequinum group (sensu Tate and Archbold 1939).

We have not examined in depth the taxonomic relationships between all the forms of $R$. megaphyllus (sensu Hill, 1992). Our conclusions, then, with respect to the specific status of the forms, $R$. simplex (simplex, keyensis, parous and


Figure 11 Plot of forearm length versus pes length for $R$. simplex subspecies, R. megaphyllus ( $\square$ ) and R. borneensis. Other taxa codes as for Figure 1.

Table 8 Mean and range, in mm, and sample size of baculum: greatest length, basal height and basal breadth, for Rhinolophus simplex subspecies and R. megaphyllus.

|  | Greatest Length | Basal Height | Basal Breadth | N |
| :--- | :--- | :--- | :--- | :--- |
| R. s. simplex | $3.00(2.67-3.48)$ | $0.85(0.67-0.99)$ | $0.84(0.60-0.95)$ | 11 |
| R. s. parvus | $2.98(2.48-3.32)$ | $0.77(0.76-0.94)$ | $0.78(0.63-0.97)$ | 3 |
| R. s. amiri subsp. nov. | $2.66(2.64-2.68)$ | $0.74(0.67-0.85)$ | $0.74(0.67-0.83)$ | 4 |
| R. megaphyllus | $2.70(2.68-2.72)$ | $0.82(0.82-0.82)$ | $0.83(0.78-0.88)$ | 2 |

amiri), are tentative. Clearly these four forms differ from $R$. megaphyllus ignifer in general size, aspects of the basicranium and shape of basal part of baculum and for parous and amiri also in the shape of the rostrum. Some independent support for this decision comes from an electrophoretic study incorporating liver tissue from specimens used in this paper, which shows that Queensland $R$. megaphyllus differs at 17 percent of their electrophoretic loci, including 4 fixed differences, from R. simplex (T. Reardon pers. comm.) However, for a complete appraisal of the relationship of these forms with R. megaphyllus it would be necessary to compare them in detail with R. m. megaphyllus and the smaller forms (R. m. vandeuseni) from the islands off northeast New Guinea. These smaller forms of R. megaphyllus, which appear to be connected to R. megaphyllus ignifer by intermediates (Koopman 1982), are approximately the same size as $R$. s. simplex (see Koopman 1982).
The taxonomic status of the Maluku form $R$. annectens (Wetar I.) is indeterminate. This form is known only from the holotype which we were unable to examine. However, this holotype was examined by J.E. Hill (pers. comm.) who stated that it is a smashed skull, which on size could be parvus, or close to it.
Prior to our study, Rhinolophus simplex was reported only from Lombok, Sumbawa and Komodo islands, (as R. s. simplex); Kai (as R. s. keyensis) and Timor (as R. celebensis parvus). We have recorded it additionally from Bali, Nusa Penida, Moyo, Sangeang, Rinca, Flores, Lembata, Alor, Sumba, Savu, Roti and Semau. Frequently on these islands it was collected from tunnels built by the Japanese during the Second World War. It was often the only species present in such tunnels.
The occurrence of morphological variation among Rhinolophus simplex from some of the Gondwanic islands of the outer Banda Arc (Sumba, Savu, Roti, Semau, Timor and Kai Kecil) reflects similar variation in the microchiropterans Hipposideros sumbae and Taphozous achates that have been examined from this region (Kitchener and Maryanto 1993; Kitchener et al. 1993). The presence on Semau of R. s. amiri, a population separated by a narrow water gap of only about three kilometres from R. s. parous on Timor, suggests reduced gene
flow is probably operating between these two populations to maintain these morphological distinctions.

## SPECIMENS EXAMINED

## Rhinolophus borneensis importunus

## INDONESIA

Java I: Kiskenda, $7^{\circ} 6^{\prime} \mathrm{S}, 110^{\circ} 16^{\prime} \mathrm{E}$, WAM (M3931013, M39319-21, M39328, M39354-5, M39361-2,


## Rhinolophus megaphyllus ignifer

## AUSTRALIA

Queensland: McIlwraith Range, $13^{\circ} 47^{\prime} \mathrm{S}, 142^{\circ} 15^{\prime} \mathrm{E}$, WAM M29972 (1 $\delta^{\circ}$ ); Iron Range, $11^{\circ} 37^{\prime} \mathrm{S}, 142^{\circ} 55^{\prime} \mathrm{E}$, WAM M29977-86, ( $60^{\circ} \delta^{\circ}, 49$ ) ); Chillagoe, $17^{\circ} 9$ 'S, $144^{\circ} 31^{\prime} \mathrm{E}$, WAM M29973 (19); Yarramulla Lava Tunnels, $18^{\circ} 13^{\prime} 30^{\prime \prime} \mathrm{S}, 144^{\circ} 40^{\prime} 30^{\prime \prime} \mathrm{E}$, WAM M29974-6 (39 \%).

## Rhinolophus simplex amiri subsp. nov (paratypes)

## INDONESIA

Savu I: Desa Menia, $10^{\circ} 29^{\prime} \mathrm{S}, 121^{\circ} 55^{\prime} \mathrm{E}$, WAM (M35113, M35117-8, M35120-25, M35127, M35129, M35132-8, M35222, 35260-2) ( $80^{\circ} \delta^{\circ}, 15$ ㅇ q).
Roti I: Baa, $10^{\circ} 44^{\prime} \mathrm{S}, 123^{\circ} 6^{\prime}$ E, WAM (M35351-2, M35370-4, M35376-8, M35380, M35389, M35391-3 ( $7 \delta^{\circ} \delta^{\circ}, 8$ 号 9 ); Sanggoen, $10^{\circ} 43^{\prime} \mathrm{S}, 123^{\circ} 9^{\prime} \mathrm{E}$, WAM M35422-3 (2 す $^{\circ}$ ).
Semau I: Uiasa, $10^{\circ} 10^{\prime} \mathrm{S}, 123^{\circ} 28^{\prime} \mathrm{E}$, WAM (M35599, M35604, M35606 (1 $\delta^{*}, 29 \%$ ); Onansila, $10^{\circ} 13^{\prime} \mathrm{S}, 123^{\circ} 30^{\circ}$ E, WAM M38014 ( $1 \delta^{\circ}$ ).

## Rhinolophus simplex parvus

## INDONESIA

Timor I: Baumata, $10^{\circ} 11^{\prime} \mathrm{S}, 123^{\circ} 43^{\prime} \mathrm{E}$, WAM (M30059, M30096-7, M30123, M30125-6, M3012840, M30145-7, M30150-2, M30155-7, WAM 30160-
 WAM (M34896, M34897-9, M34960, M34962,


Rhinolophus simplex simplex

## INDONESIA

Bali I.: Candi Kuning, $8^{\circ} 7^{\prime} \mathrm{S}, 115^{\circ} 9^{\prime} \mathrm{E}$, WAM 38441
（19）；Payongan $8^{\circ} 29^{\prime} \mathrm{E}, 115^{\circ} 15^{\prime} \mathrm{E}$ ，WAM M38424 （19）；Ubud， $8^{\circ} 30^{\prime} \mathrm{S}, 115^{\circ} 16^{\circ} \mathrm{E}$ ，WAM（M38372， M38409）（ $1 \delta^{\circ}, 1$ ）；Gianyar， $8^{\circ} 23^{\prime} \mathrm{S}, 115^{\circ} 23^{\prime} \mathrm{E}$, WAM （M38252－3，M38263，M38265，M38268，M38270，

Nusa Penida I：Sampalan， $8^{\circ} 41^{\prime} \mathrm{S}, 115^{\circ} 34^{\prime} \mathrm{E}, \mathrm{WAM}$ （M39580－1，M39584）（3q 9 ）．

Lombok I：Ngaln， $8^{\circ} 55^{\prime} \mathrm{S}, 116^{\circ} 17^{\prime} \mathrm{E}$, WAM （M31111，M33860－4）（ $2 \delta^{\circ} \delta, 4$ ㅇㅇ）．
Sumbawa I：Desa Belo， $8^{\circ} 52^{\prime}$ S， $116^{\circ} 50^{\prime} \mathrm{E}$ ，WAM （M31336－7）（1才，19）；Desa Sangeang， $8^{\circ} 18^{\prime} \mathrm{S}$ ， $118^{\circ} 56^{\prime} \mathrm{E}$ ，WAM（M31601－4，M31619）（1 $\delta^{\circ} .49$ ）．
Moyo I：Brang Kua， $8^{\circ} 14^{\prime} 15^{\prime \prime} \mathrm{S}, 117^{\circ} 36^{\prime} 45^{\prime \prime} \mathrm{E}$, WAM （M31912－5，M31921（3ठす．2 ）；Tanjung Pasir， $8^{\circ} 23^{\prime} 15^{\prime \prime} \mathrm{S}, 117^{\circ} 31^{\prime} 30^{\prime \prime} \mathrm{E}$ ，WAM（M31952－3，WAM M31962－4，M31966，M31968－79）（7o ठ̊，12와）．
Sangeang I： $8^{\circ} 13^{\prime} 30^{\prime \prime} \mathrm{S}, 119^{\circ} 00^{\prime} 20^{\prime \prime} \mathrm{E}$ ，WAM M31588 （10）．

Rinca I： $8^{\circ} 39^{\circ} \mathrm{S}, 119^{\circ} 40^{\circ} \mathrm{E}$, WAM（M32930－2， M32937－9（60 す す）．

Flores I：Daraloeng Baru， $8^{\circ} 33^{\prime} \mathrm{S}, 122^{\circ} 39^{\prime} \mathrm{E}$ ，WAM （M32589－90，M32597－8）（ 2 すす， 2 名 9 ）．

Alor I：Kalahabi， $8^{\circ} 14^{\prime} \mathrm{S}, 124^{\circ} 32^{\prime} \mathrm{E}$ ，WAM （M37615－7，M37651－2，M37654）（1才，5i ㅇ）．

Lembata I：Kampung Merdeka， $8^{\circ} 22^{\prime} \mathrm{S}, 123^{\circ} 31^{\prime} \mathrm{E}$ ， WAM M32286（19）；Desa Boto， $8^{\circ} 31^{\prime} \mathrm{S}, 123^{\circ} 23^{\prime} \mathrm{E}$ ， WAM M32429－30（ 2 ơ ${ }^{\text {o }}$ ）．

Sumba I：Waingapu， $9^{\circ} 37^{\prime} \mathrm{S}, 120^{\circ} 14^{\prime} \mathrm{E}$ ，WAM （M30249－50，M30252－3）（3 ठ̊，1\％）．Bondokodi $9^{\circ} 35^{\circ} \mathrm{S}, 119^{\circ} 8^{\prime} \mathrm{E}$ ，WAM（M30486，M30492）（ 2 o $^{\star}$ ठ ） ．

## Rhinolophus simplex keyensis

Kai Kecil I：Tual， $5^{\circ} 38^{\prime} \mathrm{S}, 132^{\circ} 44^{\prime} \mathrm{E}$ ，WAM （M42642－3（1才，19）．

## ACKNOWLEDGEMENTS

We gratefully acknowledge the support of Mr J． Bannister，Director，Western Australian Museum during this research（now retired）；and Dr M． Amir，Director，Balitbang Zoologi（LIPI）．The Directors of the Indonesian Department responsible for the conservation of wildlife in Nusa Tenggara（BKSDA），Ir．J．Mochtar（NTT）and Ir．P． Supriadi（NTB）and Maluku Tenggara，Ir．J． Rustandi，provided us with great assistance in the field，as did a number of their staff．

To our colleagues who participated in the field work，particularly Dr R．A．How and Mr R．E． Johnstone，Western Australian Museum；Bapak Boeadi，Ir Maharadatunkamsi，and Ir I．Maryanto， Museum Zoologicum Bogoriense，we extend our thanks for their effort and companionship．

We are particularly grateful to Ms Sue Hisheh and Mr Paul Ottaviana for assistance with the protein electrophoresis．
Expedition costs were defrayed by grants to Dr D．J．Kitchener from：National Geographic Society， Washington；Australian National Parks and

Wildlife Service，Canberra，and The Mark Mitchell Trust，South Australia；and a grant to Drs L．H． Schmitt，D．J．Kitchener and R．A．How from the Australian Research Council，Canberra．Garuda Indonesia kindly defrayed freight costs of the expeditions．
Mrs N．Cooper，Western Australian Museum，ran the computer analyses．Mrs S．Dalton and Mrs Anne Nevin，Western Australian Museum，typed the manuscript．
We are particularly grateful to Mr J．E．Hill，The Natural History Museum，London，and Dr R．A． How，Western Australian Museum，who critically commented on an earlier draft of the MS．

## REFERENCES

Andersen，K．（1905）．On some bats of the genus Rhinolophus，with some remarks on their mutual affinities，and descriptions of twenty－six new forms． Proceedings of the Zoological Society of London 2：75－ 145.

Andersen，K．（1918）．Diagnoses of new bats of the families Rhinolophidae and Megadermatidae．Annals and Magazine of Natural History 2：374－384．
Bergmans，W．and van Bree，P．J．H．（1986）．On a collection of bats and rats from the Kangean islands， Indonesia（Mammalia ：Chiroptera and Rodentia）． Zeitschrift Saügetierk 51：329－344．
Corbet，G．B．and J．E．Hill（1980）．A world list of mammalian species．British Museum（Natural History）， London．
Genstat 5 Committee（1987）．Genstat 5 Reference Manual． Oxford University Press，Oxford．
Goodwin，R．E．（1979）．The bats of Timor：systematics and ecology．Bulletin of the American Museum of Natural History．163：73－122．
Gray，J．E．（1834）．Characters of a new species of bat （Rhinolophus，Geoffry）from New Holland． Proceedings of the Zoological Society of London 52－34．
Hill，J．E．（1983）．Bats（Mammalia：Chiroptera）from Indo－ Australia．Bulletin of the British Museum of Natural History（Zoology）45：103－208．
Hill，J．E．（1992）．Chiroptera．In Mammals of the IndoMalayan Region：a systematic review，pp 54－161， Natural History Museum Publ．and Oxford University Press，Oxford．
Hill，J．E．and Smith，J．D．（1984）．Bats，a natural history． British Museum（Natural History），London．
Honacki，J．H．，Kinman，K．E．and Koeppl，J．W．（1982）． Mammal species of the world．A taxonomic and geographic reference．Allen Press，Lawrence，Kansas．
Kitchener，D．J．and Maryanto，I．（1993）．Taxonomic reappraisal of the Hipposideros larvatus species complex（Chiroptera ：Hipposideridae）in the Greater and Lesser Sunda islands，Indonesia．Records of the Western Australian Museum 16：119－173．
Kitchener，D．J．，Schmitt，L．H．，Hisheh，S．，How，R．A．， Cooper，N．K．and Maharadatunkamsi（1993）．

Morphological and genetic variation in the Bearded Tomb Bats (Taphozous : Emballonuridae) of Nusa Tenggara, Indonesia. Mammalia 57: 63-83.
Koopman, K.F. (1982). Results of the Archbold Expeditions No. 109. Bats from eastern Papua and the eastern Papuan islands. American Museum Novitates 2747: 1-34.
Nei, M. (1978). Estimation of average heterozygosity and genetic distance from a small number of individuals. Genetics 89: 583-590.
Nevo, E., Beiles, A. and Ben-Shlomo, R. (1984). The evolutionary significance of genetic diversity: ecological, demographic and life history correlates. In G.S. Mani (ed.) Evolutionary dynamics of genetic diversity, pp. 13-213. Springer-Verlag, Heidelberg.
Peters, W. (1871). Uber die Gattungen und Arten der Hufeisennasen, Rhinolophi. Monatsbericht der Kgl. Preussischen Akademie der Wissenschaften 302-332.
Richardson, B.J., Baverstock, P.R. and Adams, M. (1986). Allozyme electrophoresis: a handbook for animal systematics and population studies. Academic Press, Sydney.
Sanborn, C.C. (1939). Eight new bats of the genus

Rhinolophus. Publications of the Field Museum of natural History, Zoology Series 24: 37-43.
Smithe, F.B. (1975). Naturalist's color guide. American Museum of Natural History, New York.
Sneath, P.H.A. and Sokal, R.R. (1973). Numerical Taxonomy. W.H. Freeman, San Francisco.
Swofford, D.L. and Selander, R.B. (1989). BIOSYS-1: a computer program for the analysis of allelic variation in population genetics and biochemical systematics. Release 1.7.

Tate, G.H.H. and Archbold, R. (1939). Results of the Archbold Expeditions No. 24. Oriental Rhinolophus with special reference to material from the Archbold collections. American Museum Novitates 1036: 1-14.
van Strien, N.J. (1986). Abbreviated checklist of the mammals of the Australasian Archipelago. School of Environmental Conservation Management, Bogor, Indonesia.
Weir, B.S. and Cockerham, C.C. (1984). Estimating Fstatistics for the analysis of population structure. Evolution 38: 1358-1370.

Manuscript received 18 August 1993; accepted 1 March 1994.

